Terpenoids and the mechanism of their anticancer effects

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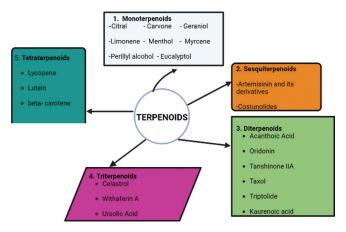
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Abstract

Terpenoids otherwise called isoprenoids are secondary metabolites with diverse biological and pharmacological activities. They serve as candidate compounds for drug discovery. They are classified into mono-, sesqui-, di-, tri-, tetra-, and polyterpenoids based on their isoprenoid structural units. They exhibit anticancer properties through various mechanisms such as cell cycle arrest induction, angiogenesis suppression, decreased tumor cell differentiation, and apoptosis. Most terpenoids used in cancer therapy are derived from medicinal plants. This paper reviews important plant-derived terpenoids used in cancer treatment, their medicinal plant sources, and their mechanism of action.

Key words: Cancer, plants, terpenoids

GRAPHICAL ABSTRACT



INTRODUCTION

ancer is a global complex disease characterized by sustaining proliferative signaling, evading growth suppressors, enabling replicative immortality, resisting death, inducing angiogenesis, and activating invasion and metastasis, apart from reprogramming energy metabolism and evading immune destruction.[1] It is a worldwide health and economic concern for both developed and developing countries. It is the leading cause of global deaths. According to the World Health Organization, over 10 million people died of cancer in 2020.[2] Breast and prostate cancers have been recorded to have the highest incidence, prevalence, and mortality rates in females and males, respectively. Most cancer death rates increase as a result of late diagnosis,^[3] while early detection and treatment can reduce morbidity and mortality rates.

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Surgery, chemotherapy, and radiotherapy stem cell renewal are the major effective approaches in cancer therapy. [4] However, treatment failures, drug resistance, and side effects are the major drawbacks of cancer chemotherapy. This has necessitated the search for other alternative novel drugs with better efficacy and fewer side effects. Plant diet has also been recorded to protect the human body from the risk of carcinogenesis. [5] Plants contain phytochemicals and their derivatives with varying pharmacological activities. Over the decades, plants' natural products have been documented as an important source of drug discovery. At present, plant-derived drugs are becoming major sources of cancer treatment. [6,7] This review aims at documenting information on some plant-based terpenoids and their mechanism of anticancer activity.

TERPENOIDS

Terpenoids also called isoprenoids are modified terpenes that contain different functional groups and an oxidized methyl group. Terpenoids are ubiquitously distributed in nature, the most diverse group of secondary metabolites, and present in plants and lower invertebrates. They are rich sources of candidate compounds for drug discovery. They are obtained from mevalonic acid (MVA) which consists of isoprene (C_5) structural units. Most terpenoids are isolated from medicinal plants that belong to the following plant groups: Oleaceae, Rutaceae, Labiatae, Acanthaceae, Compositae, Taxaceae, Pinaceae, Lauraceae, Araliaceae, Celastraceae, etc. Terpenoids are classified according to the number of isoprene (2-methylbuta-1, 3-diene) units into monoterpenoids (C_{10}),

Table 1: Terpenoid classification and sources			
Terpenoid subclass	Compounds	Sources	References
Monoterpenoids	Citral	Lemon grass oil (Cymbopogon citratus)	[12]
	Carvone	Spearmint oil (<i>Mentha spicata</i>) and Caraway oil (<i>Carum carvi</i>),	[13]
	Geraniol	Palmarosa oil, rose oil, and ninde oil (<i>Aeollanthus myrianthus</i>)	[14,15]
	Limonene	Citrus essential oils: orange (Citrus sinensis), lime (Citrus aurantifolia), grape (Citrus paradisi), mandarin (Citrus reticulata), and lemon (Citrus limon)	[16]
	Menthol	Peppermint oil (Mentha piperita)	[17]
	Myrcene	bay laurel oil (<i>Laurus nobilis</i>) and Verbena oil (<i>Lippia citriodora</i>)	[18]
	Perillyl alcohol		[19]
	Eucalyptol (1.8-cineole)	Eucalyptus leaf oil (Eucalyptus globules), rosemary (Rosmarinus officinalis)	[20]
Sesquiterpenoids	Artemisinin and its derivatives	Sweet wormwood (Artemisia annua)	[21]
	Costunolide	Aucklandia lappa	[22,23]
Diterpenoids	Acanthoic acid	Acanthopanax koreanum Nakai, Croton oblongifolius Roxb	[24,25]
	Oridonin	Rabdosia rubescens	[26]
	Tanshinone IIA	Salvia miltiorrhiza Bge	[27]
	Taxol (Paclitaxel)	Taxus brevifolia	[28]
	Triptolide	Tripterygium wilfordii	[29,30]
	Kaurenoic acid	Annona senegalensis (Annonaceae)	[31]
Triterpenoids	Celastrol	Tripterygium wilfordii	[29]
	Withaferin A	Withania somnifera	[32]
	Ursolic acid	Rosmarinus officinalis (rosemary), Calluna vulgaris and Eugenia jambolana, Salvia officinalis, and Eriobotrya japonica	[33]
Tetraterpenoids	Lycopene	Tomatoes (<i>Lycopersicon esculentum</i>), pink guavas, apricots, water melon, and pink grapefruits	[34,35]
	Lutein	Spinach, kale, carrot, corn, and egg yolk	[36,37]
	β -carotene	Carrot	[38]

sesquiterpenoids (C_{15}), diterpenoids (C_{20}), triterpenoids (C_{30}), tetraterpenoids (C_{40}), and polyterpenoids (C > 40) [Table 1]. They possess vast medicinal importance that includes: antitumor, antibacterial, antimalarial, anti-inflammatory, antiviral, anticancer, immunomodulatory, anti-aging, antioxidant, anti-depressants, antifungal, and antidiabetic. [8,9]

Mevalonate (MVA) and 2C-methyl-D-erythritol-4- phosphate (MEP) pathways are the two major pathways for terpenoid biosynthesis. The main metabolic intermediate for both pathways is isopentenyl diphosphate (1PP). Plants produce monoterpenoids, diterpenoids, and tetraterpenoids through the MEP in the plastids, whereas sesquiterpenoids and triterpenoids are produced through the mevalonate route in the cytoplasm.

Terpenoids of natural origin are known to possess anticancer properties. A huge number of triterpenoids have been known to subdue the growth of cancer cells without producing toxicity in normal cells.^[10] The major mechanisms of anticancer activities of terpenoids include induction of cell cycle arrest, angiogenesis suppression, decreased tumor cell differentiation, apoptosis, anti-proliferative, and anti-angiogenic.^[5,11]



Isoprene moiety

MONOTERPENOIDS

Citral

It is effective against P388 mouse leukemia, HeLa and ECC-1 cancer cells.^[39] It also induces apoptosis driving lipogenesis in both *in vitro* and *in silico* studies. It alters the potential of the mitochondrial membrane, increases intracellular reactive oxygen species, and initiates the death of cancer cells by apoptosis.^[40]

Carvone

It is a cytotoxic agent against HeLa cells.^[1] It is known to increase intracellular reactive oxygen species mediated apoptotic cell death in cancer cells.^[41]

Geraniol

It blocks tumor cell growth by inhibiting the G_1 phase Michigan Cancer Foundation-7 (MCF-7) breast cancer cell cycle. [14] It also inhibits the growth of HepG2 human hepatic carcinoma cells by reducing the activity of 3-hydroxymethylglutaryl coenzyme A reductase which results in a decrease in cancer growth and cholesterol biosynthesis. [1] In addition, it induces apoptosis and elevates Bak expression, a proapoptotic protein, in cultured pancreatic tumor cells [42] and induces angiogenesis. [43] Duncan

et al. reported that geraniol reduces cell division, cell cycle progression, and cyclin-dependent kinase 2 activity in MCF-7 breast cancer cells, claim Duncan *et al.*^[44]

Limonene

D-limonene antiangiogenic, proapoptotic, and anti-oxidant actions reduce the rate of tumor growth and metastasis.^[45,46]

Menthol

It is a concentration-dependent cytotoxic agent against murine leukemia WEHI-3 cells.^[47] Menthol decreased the proliferation and motility of prostate cancer DU145 cells, according to research by Wang *et al*.^[48]

Myrcene

It is known to be cytotoxic against MCF-7 breast carcinoma, gall tumor, human colon adenocarcinoma (HT-29), and HeLa (Human Cervical carcinoma). [49] It induces oxidative stress and apoptosis in cancer cells. [50]

Perillyl Alcohol

It is a monoterpenoid derived from lavender, peppermint, and other plants that block telomerase activity in prostate cancer.^[51]

SESQUITERPENOIDS

Artemisinin and Derivatives

Artemisinin and its derivatives are globally known as an antimalarial agent. The anticancer properties of artemisinin are mediated through the induction of cell cycle arrest, boosting ferroptosis and autophagy, inducing cell death, limiting cell metastasis, and suppressing cancer growth. [52-54] It is active against colorectal, cervical, hepatocellular, leukemia, breast, prostate, colon, gastric, melanoma, and lung cancer. [55,56]

Costunolide

It is a sesquiterpene lactone compound obtained from a medicinal plant *Aucklandia lappa* Decne. It manifests anticancer activity by inhibiting cancer cell proliferation, inhibiting metastasis, inhibiting angiogenesis, inducing cancer cell apoptosis and differentiation, inhibiting cell cycle progression, and reversing multidrug resistance.^[57,58]

Oridonin

It is a diterpenoid that induces apoptosis in lung and breast cancer cells through the arrest of G2/M cell cycle progression,

resulting to the inhibition of cancer progression. It also facilitates the phagocytosis of apoptotic tumor cells through the regulation of macrophage functioning that involves tumor necrosis factor (TNF)- α and interleukin-1 β . [27,59,60] It is effective against breast, colon, pancreatic, lung, gastric, prostate, and skin cancers. [61]

Tanshinone IIA

Its antitumor/cancer activity is attributed to its tumor growth inhibition, apoptosis induction, signaling pathway, and cell cycle regulation. [62,63] It also inhibits angiogenesis and it is active against breast cancers, [64] and leukemia, [65] lung cancer, [66,67] gastric carcinomas, [68-70] colorectal cancer, [71,72] glioma, [73] osteosarcoma, [74] cervical cancer, [75,76] ovarian cancer, [77] and prostate cancer. [78]

Triptolide

It is a diterpenoid triepoxide isolated from *Tripterygium wilfordii* Hook. All 60 cancer cell lines from the US National Cancer Institute are inhibited from proliferating by triptolide. In addition, by causing the biggest RNA polymerase II (Rpb1) subunit of cancer cells to degrade in a proteasome-dependent manner, it hinders global gene transcription.^[79]

Paclitaxel

It is a well-known anti-neoplastic substance that was originally discovered in the bark of pacific yew trees (*Taxus brevifolia*). Its anticancer action is by inducting mitotic arrest through the targeting of the cytoskeleton component tubulin, resulting in mitotic activation and apoptosis.^[30] It is used to treat pancreatic, ovarian, and breast cancers.

TRITERPENOIDS

Celastrol

A quinone-methide triterpene called celastrol is derived from the plant lei gong teng (*T. wilfordii Hook F*). It stops cancer growth by initiating TNF-α induced NF-kB signaling pathway.^[80] In addition, it prevents the growth, migration, and invasion of chondrosarcoma cells by inhibiting the protein phosphatase 2A-Akt signaling pathway *in vivo*.^[81] Celastrol activates cell cycle arrest and human cancer cell death through apoptosis.^[82]

Withaferin A (WA)

WA is obtained from the medicinal plant *Withania somnifera* Dunal (Ashwagandha). It exerts its anticancer activity by activating apoptosis and G2/M cell cycle arrest.^[83]

Ursolic Acid (UA)

It is an apentacyclic triterpene acid that is present in the leaves of Ocimum as well the berries, leaves, flowers, and fruits of *Eriobotrya japonica*, *Calluna vulgaris*, *Eugenia jambolana*, *Rosmarinus officinalis*, and *Salvia officinalis*.^[33] Ursolic acid inhibits the development and spread of cancer by reducing proliferation and triggering apoptosis both *in vitro* and *in vivo*. Furthermore, it dose-dependently prevents the *in vitro* growth of the gastric cancer cell line BGC-803. It also prevents breast cancer multiplication by inducing G1/G2 cell arrest and regulates the expression of key proteins in signal transduction pathways.^[84]

TETRATERPENOIDS

The most prevalent terpenoids are tetraterpenoids.

Lutein

According to Li *et al.*, lutein suppresses hypoxia-induced proliferation, invasion, and migration of cancer cells in the breast. [85]

Lycopene

It is the main pigment found in tomatoes. Lycopene is known to prevent cell proliferation, induce apoptosis, inhibit cell invasion, and inhibit cell cycle progression, angiogenesis, and metastasis.^[86]

Beta-carotene (β-carotene)

Beta-carotene can also be referred to as provitamin A. Beta-carotenes are known to induce apoptosis, cell cycle arrest, signaling pathways, migration invasion, and metastasis.^[87]

CONCLUSION

The knowledge of ethnobotanicals is one of the major approaches to new drug discovery. Plants provide the major sources for modern anticancer drug discovery. Numerous terpenoid medications have produced significant therapeutic and commercial benefits. Terpenoids have, over the years, continued to be indispensable in drug discovery. Terpenoids that are naturally produced have opened up new prospects for researchers to identify and use novel compounds against cancer and other diseases with minimal side effects.

The present review summarizes the basic natural terpenoids currently used in the treatment of cancer. The review also highlighted the plants responsible for the anticancer activity as well as their corresponding pharmacological actions.

REFERENCES

- Sobral MV, Xavier AL, Lima TC, de Sousa DP. Antitumor activity of monoterpenes found in essential oils. ScientificWorldJournal 2014;2014:953451.
- Ma L, Zhang M, Zhao R, Wang D, Ma Y, Li A. Plant natural products: Promising resources for cancer chemoprevention. Molecules 2021;26:933.
- Miller KD, Nogueira L, Mariotto AB, Rowland JH, Yabroff KR, Alfano CM, et al. Cancer treatment and survivorship statistics, 2019. CA Cancer J Clin 2019;69:363-85.
- 4. Haider T, Pandey V, Banjare N, Gupta PN, Soni V. Drug resistance in cancer: Mechanisms and tackling strategies. Pharmacol Rep 2020;72:1125-51.
- Rahman MA, Bulbul MR, Kabir Y. Plant-based products in cancer prevention and treatment. In: Functional Foods in Cancer Prevention and Therapy. Cambridge: Academic Press; 2020. p. 237-59.
- Tilaoui M, Mouse HA, Zyad A. Update and new insights on future cancer drug candidates from plant-based alkaloids. Front Pharmacol 2021;12:719694.
- 7. Bergman ME, Davis B, Philips MA. Medically useful plant terpenoids: Biosynthesis, occurrence, and mechanism of action. Molecules 2019;24:3961.
- 8. Cox-Georgian D, Ramadoss N, Dona C, Basu C. Therapeutic and medicinal uses of terpenes. In: Medicinal Plants. Vol. 12. Germany: Springer; 2019. p. 333-59.
- 9. Yang W, Chen X, Li Y, Guo S, Wang Z, Yu X. Advances in pharmacological activities of terpenoids. Nat Prod Commun 2020;15:1-13.
- Ansari IA, Akhtar MS. Current insights on the role of terpenoids as anticancer agents: A perspective on cancer prevention and treatment. In: Swamy M, Akhtar M, editors. Natural Bio-active Compounds. Singapore: Springer; 2020. p. 53-80.
- 11. Chopra B, Dhingra AK, Dhar KL, Nepali K. Emerging role of terpenoids for the treatment of cancer: A review. Mini Rev Med Chem 2021;21:2300-36.
- 12. Balusamy SR, Perumalsamy H, Veerappan K, Huq MA, Rajeshkumar S, Lakshmi T, et al. Citral induced apoptosis through modulation of key genes involved in fatty acid biosynthesis in human prostate cancer cells: *In silico* and in vitro study. Biomed Res Int 2020;2020:6040727.
- 13. Yoshida E, Kojima M, Suzuki M, Matsuda F, Shimbo K, Onuki A, et al. Increased carvone production in Escherichia coli by balancing limonene conversion enzyme expression via targeted quantification concatamer proteome analysis. Sci Rep 2021;11:22126.
- 14. Baser KH, Kurkcuoglu M, Demirci B. Ninde oil (*Aeollanthus myrianthus* Taylor) revisited: Analysis of a historical oil. J Essent Oil Res 2005;17:137-8.
- 15. Chen W, Viljoen AM. Gernaniol-a review of a commercially important fragrance material. S Afr J Bot 2010;76:643-51.
- 16. Schween JH, Dlugi R, Hewitt CN, Foster P. Determination and accuracy of VOC-fluxes above the pine/oak forest at Castelporziano. Atmos Environ 1997;31:199-215.

- 17. Croteau RB, Davis EM, Ringer KL, Wildung MR. (-)-Menthol biosynthesis and molecular genetics. Naturwissenschaften 2005;92:562-77.
- 18. Santos PM, Sa-Correia I. Adaptation to beta-myrcene catabolism in *Pseudomonas* sp. M1: An expression proteomics analysis. Proteomics 2009;9:5101-11.
- 19. Belanger JT. Perillyl alcohol: Applications in oncology. Altern Med Rev 1998;3:448-57.
- 20. Izham MN, Hussin Y, Rahim NF, Aziz MN, Yeap SK, Rahman HS, *et al.* Physicochemical characterization, cytotoxic effect and toxicity evaluation of nanostructured lipid carrier loaded with eucalyptol. BMC Complement Med Ther 2021;21:254.
- 21. Anibogwu R, De Jesus K, Pradhan S, Pashikanti S, Mateen S, Sharma K. Extraction, isolation and characterization of bioactive compounds from artemisia and their biological significance: A review. Molecules 2021;26:6995.
- 22. Yang YI, Kim JH, Lee KT, Choi JH. Costunolide induces apoptosis in platinum-resistant human ovarian cancer cells by generating reactive oxygen species. Gynecol Oncol 2011;123:588-96.
- 23. Liu XN, Li HM, Wang SP, Zhang JZ, Liu DL. Sesquiterpene lactones of *Aucklandia lappa*: Pharmacology, pharmacokinetics, toxicity, and structure-activity relationship. Chin Herb Med 2021;13:167-76.
- 24. Jung MG, Do GM, Shin JH, Ham YM, Park SY, Kwon O. Acanthopanax koreanum Nakai modulates the immune response by inhibiting TLR 4-dependent cytokine production in rat model of endotoxic shock. Nutr Res Pract 2013;7:460-5.
- Sittithumcharee G, Kariya R, Kasemsuk T, Saeeng R, Okada S. Antitumor effect of acanthoic acid against primary effusion lymphoma via inhibition of c-FLIP. Phytother Res 2021;35:7018-26.
- 26. Guan YF, Liu XJ, Pang XJ, Liu WB, Yu GX, Li YR, *et al.* Recent progress of oridonin and its derivatives for cancer therapy and drug resistance. Med Chem Res 2021;30:1795-821.
- 27. Pan Y, Chen L, Li R, Liu Y, Nan M, Hou L. Tanshinone IIa induces autophagy and apoptosis via PI3K/Akt/mTOR Axis in acute promyelocytic leukemia NB4 cells. Evid Based Complement Alternat Med 2021;2021:3372403.
- 28. Weaver BA. How Taxol/paclitaxel kills cancer cells. Mol Biol Cell 2014;25:2677-81.
- 29. Huang M, Lu JJ, Huang MQ, Bao JL, Chen XP, Wang YT. Terpenoids: Natural products for cancer therapy. Expert Opin Investig Drugs 2012;21:1801-18.
- Seo EJ, Dawood M, Hult AK, Olsson ML, Efferth T. Network pharmacology of triptolide in cancer cells: Implications for transcription factor binding. Invest New Drugs 2021;39:1523-37.
- 31. Okoye TC, Akah PA, Nworu CS, Ezike AC. Kaurenoic acid isolated from the root bark of *Annona senegalensis* induces cytotoxic and antiproliferative effects against PANC-1 and HeLa cells. Eur J Med Plants 2014;4:579-89.
- 32. Sivasankarapillai VS, Nair RK, Rahdar A, Bungau S, Zaha DC, Aleya L, *et al.* Overview of the anticancer

- activity of withaferin A, an active constituent of the Indian ginseng *Withania somnifera*. Environ Sci Pollut Res Int 2020;27:26025-35.
- 33. Liu J. Pharmacology of oleanolic acid and ursolic acid. J Ethnopharmacol 1995;49:57-68.
- 34. Puah BP, Jalil J, Attiq A, Kamisah Y. New insights into molecular mechanism behind anti-cancer activities of lycopene. Molecules 2021;26:3888.
- 35. Khan UM, Sevindik M, Zarrabi A, Nami M, Ozdemir B, Kaplan DN, *et al.* Lycopene: Food sources, biological activities, and human health benefits. Oxid Med Cell Longev 2021;2021:2713511.
- 36. Krinsky NI, Landrum JT, Bone RA. Biologic mechanisms of the protective role of lutein and zeaxanthin in the eye. Ann Rev Nutr 2003;23:171-201.
- Vasanthkumar T, Hanumanthappa M, Prabhakar B, Hanumanthappa SK. Hepatoprotective effect of curcumin and capsaicin against lipopolysaccharide induced liver damage in mice. Pharmacogn J 2017;9: 947-51.
- 38. Kumari S, Goyal A, Garg M. Phytochemistry and pharmacological update on tetraterpenoids. Nat Prod J 2021;11:617-28.
- 39. Xia H, Liang W, Song Q, Chen X, Chen X, Hong J. The *in vitro* study of apoptosis in NB4 cell induced by citral. Cytotechnology 2013;65:49-57.
- 40. Sanches LJ, Marinello PC, Panis C, Fagundes TR, Morgado-Díaz JA, de-Freitas-Junior JCM, et al. Cytotoxicity of citral against melanoma cells: The involvement of oxidative stress generation and cell growth protein reduction. Tumor Biol 2017;39:1010428317695914.
- 41. Iyappan P, Bala MD, Sureshkumar M, Veeraraghavan VP, Palanisamy A. D-carvone induced ROS mediated apoptotic cell death in human leukemic cell lines (Molt-4). Bioinformation 2021;17:171-80.
- 42. Burke YD, Ayoubi AS, Werner SR, McFarland BC, Heilman DK, Ruggeri BA, *et al*. Effects of the isoprenoids perillyl alcohol and farnesol on apoptosis biomarkers in pancreatic cancer chemoprevention. Anticancer Res 2002;22:3127-34.
- 43. Wittig C, Scheuer C, Parakenings J, Menger MD, Laschke MW. Geraniol suppresses angiogenesis by downregulating vascular endothelial growth factor (VEGF)/VEGFR-2 signaling. PLoS One 2015;10:e0131946.
- 44. Duncan RE, Lau D, El-Sohemy A, Archer MC. Geraniol and beta-ionone inhibit proliferation, cell cycle progression, and cyclin-dependent kinase 2 activity in MCF-7 breast cancer cells independent of effects on HMG-CoA reductase activity. Biochem Pharmacol 2004;68:1739-47.
- 45. Alipanah H, Farjam M, Zarenezhad E, Roozitalab G, Osanloo M. Chitosan nanoparticles containing limonene and limonene-rich essential oils: Potential phytotherapy agents for the treatment of melanoma and breast cancers. BMC Complement Med Ther 2021;21:186.
- 46. Lu XG, Zhan LB, Feng BA, Qu MY, Yu LH, Xie JH.

- Inhibition of growth and metastasis of human gastric cancer implanted in nude mice by d-limonene. World J Gastroenterol 2004;10:2140-4.
- 47. Lu HF, Liu JY, Hsueh SC, Yang YY, Yang JS, Tan TW, et al. (-)-Menthol inhibits WEHI-3 leukemia cells in vitro and in vivo. In Vivo 2007;21:285-9.
- 48. Wang Y, Wang X, Yang Z, Zhu G, Chen D, Meng Z. Menthol inhibits the proliferation and motility of prostate cancer DU145 cells. Pathol Oncol Res 2012;18:903-10.
- 49. Da Silva SL, Figueiredo PM, Yano T. Cytotoxic evaluation of essential oil from *Zanthoxylum rhoifolium* Lam. leaves. Acta Amazon 2007;37:281-6.
- 50. Bai X, Tang J. Myrcene exhibits antitumor activity against lung cancer cells by inducing oxidative stress and apoptosis mechanism. Nat Prod Commun 2020;15:1-7.
- 51. Sundin T, Peffley DM, Gauthier D, Hentosh P. The isoprenoid perillyl alcohol inhibits telomerase activity in prostate cancer cells. Biochimie 2012;94:2639-48.
- 52. Zhang B. Artemisinin-derived dimers as potential anticancer agents: Current developments, action mechanisms, and structure-activity relationships. Arch Pharm (Weinheim) 2020;353:e1900240.
- 53. Li D, Zhang J, Zhao X. Mechanisms and molecular targets of artemisinin in cancer treatment. Cancer Invest 2021;39:675-84.
- 54. Zhu S, Yu Q, Huo C, Li Y, He L, Ran B, *et al*. Ferroptosis: A novel mechanism of artemisinin and its derivatives in cancer therapy. Curr Med Chem 2021;28:329-34.
- 55. Babaei G, Aliarab A, Abroon S, Rasmi Y, Aziz SG. Application of sesquiterpene lactone: A new promising way for cancer therapy based on anticancer activity. Biomed Pharmacother 2018;106:239-46.
- Lee SH, Cho YC, Lim SL. Costunolide, a sesquiterpene lactone, suppresses skin cancer via induction of apoptosis and blockage of cell proliferation. Int J Mol Sci 2021;22:2075.
- 57. Okubo S, Ohta T, Fujita H, Shoyama Y, Uto T. Costunolide and dehydrocostuslactone from *Saussurea lappa* root inhibit autophagy in hepatocellular carcinoma cells. J Nat Med 2021;75:240-5.
- 58. Qi X, Zhang D, Xu X, Feng F, Ren G, Chu Q, *et al.* Oridonin nanosuspension was more effective than free oridonin on G2/M cell cycle arrest and apoptosis in the human pancreatic cancer PANC-1 cell line. Int J Nanomedicine 2012;7:1793-804.
- 59. Liu X, Xu J, Zhou J, Shen Q. Oridonin and its derivatives for cancer treatment and overcoming therapeutic resistance. Genes Dis 2021;8:448-62.
- 60. Abdullah NA, Hashim NM, Ammar A, Zakuan NM. An insight into the anti-angiogenic and anti-metastatic effects of oridonin: Current knowledge and future potential. Molecules 2021;26:775.
- 61. Kim JH, Lee JO, Lee SK, Kim N, You GY, Moon JW, et al. Celastrol suppresses breast cancer MCF-7 cell viability via the AMP-activated protein kinase (AMPK)-induced p53-polo like kinase 2 (PLK-2) pathway. Cell Signal 2013;25:805-13.

- 62. Fang ZY, Zhang M, Liu JN, Zhao X, Zhang YQ, Fang L. Tanshinone IIA: A review of its anticancer effects. Front Pharmacol 2021;11:611087.
- YuT, Zhou Z, MuY, de Lima G, Luo KQ. A novel anti-cancer agent, acetyltanshinone IIA, inhibits oestrogen receptor positive breast cancer cell growth by down-regulating the oestrogen receptor. Cancer Lett 2014;346:94-103.
- 64. Yun SM, Jung JH, Jeong SJ, Sohn EJ, Kim B, Kim SH. Tanshinone IIA induces autophagic cell death via activation of AMPK and ERK and inhibition of mTOR and p70 S6K in KBM-5 leukemia cells. Phytother Res 2013;28:458-64.
- 65. Cheng CY, Su CC. Tanshinone IIA may inhibit the growth of small cell lung cancer H146 cells by up-regulating the Bax/Bcl-2 ratio and decreasing mitochondrial membrane potential. Mol Med Rep 2010;3:645-50.
- 66. Xie J, Liu J, Liu H, Liang S, Lin M, Gu Y, *et al.* The antitumor effect of tanshinone IIA on anti-proliferation and decreasing VEGF/VEGFR2 expression on the human non-small cell lung cancer A549 cell line. Acta Pharm Sin B 2015;5:554-63.
- 67. Chen J, Shi DY, Liu SL, Zhong L. Tanshinone IIA induces growth inhibition and apoptosis in gastric cancer *in vitro* and *in vivo*. Oncol Rep 2012;27:523-8.
- 68. Su CC. Tanshinone IIA inhibits gastric carcinoma AGS cells by decreasing the protein expression of VEGFR and blocking Ras/Raf/MEK/ERK pathway. Int J Mol Med 2018;41:2389-96.
- 69. Xu Z, Chen L, Xiao Z, Zhu Y, Jiang H, Jin Y, *et al.* Potentiation of the anticancer effect of doxorubicinin drug-resistant gastric cancer cells by tanshinone IIA. Phytomedicine 2018;51:58-67.
- 70. Su CC, Lin YH. Tanshinone IIA down-regulates the protein expression of ErBb-2 and up-regulates TNF-alpha in colon cancer cells *in vitro* and *in vivo*. Int J Mol Med 2008;22:847-51.
- Ma S, Lei Y, Zhang L, Wang J. Research on the inhibiting effect of tanshinone IIA on colon cancer cell growth via COX-2-Wnt/β-catenin signaling pathway. J BUON 2018;23:1337-42.
- 72. Ding L, Wang S, Wang W, Wang W, Lv P, Zhao D, *et al.* Tanshinone IIA affects autophagy and apoptosis of glioma cells by inhibiting phosphatidylinositol 3-kinase/Akt/mammalian target of rapamycin signaling pathway. Pharmacology 2017;99:188-95.
- 73. Yen JH, Huang ST, Huang HS, Fong YC, Wu YY, Chiang JH, *et al.* HGK-sestrin 2 signaling-mediated autophagy contributes to antitumor efficacy of tanshinone IIA in human osteosarcoma cells. Cell Death Dis 2018;9:1003-17.
- 74. Pan TL, Wang PW, Hung YC, Huang CH, Rau KM. Proteomic analysis reveals tanshinone IIA enhances apoptosis of advanced cervix carcinoma CaSki cells through mitochondria intrinsic and endoplasmic reticulum stress pathways. Proteomics 2013;13:3411-23.

- 75. Qin J, Shi H, Xu Y, Zhao F, Wang Q. Tanshinone IIA inhibits cervix carcinoma stem cells migration and invasion via inhibiting YAP transcriptional activity. Biomed Pharmacother 2018;105:758-65.
- 76. Jiao JW, Wen F. Tanshinone IIA acts via p38 MAPK to induce apoptosis and the down-regulation of ERCC1 and lung-resistance protein in cisplatin-resistant ovarian cancer cells. Oncol Rep 2011;25:781-8.
- 77. Won SH, Lee HJ, Jeong SJ, Lee HJ, Lee EO, Jung DB, *et al.* Tanshinone IIA induces mitochondria dependent apoptosis in prostate cancer cells in association with an inhibition of phosphoinositide 3-kinase/AKT pathway. Biol Pharm Bull 2010;33:1828-34.
- 78. Vispé S, DeVries L, Créancier L, Besse J, Bréand S, Hobson DJ, et al. Triptolide is an inhibitor of RNA polymerase I and II-dependent transcription leading predominantly to down-regulation of short-lived mRNA. Mol Cancer Ther 2009;8:2780-90.
- Wang Y, Lu JJ, He L, Yu Q. Triptolide (TPL) inhibits global transcription by inducing proteasome-dependent degradation of RNA polymerase II (Pol II). PLoS One 2011;6:e23993.
- 80. Kim JY, Song JJ, Kwo BM, Lee JD. Tanshinone IIA exerts antitumor activity against vestibular schwannoma cells by inhibiting the expression of hypoxia-inducible factor-1α. Mol Med Rep 2015;12:4604-9.
- 81. Liu Z, Ma L, Wen ZS, Hu Z, Wu FQ, Li W, *et al.* Cancerous inhibitor of PP2A is targeted by natural compound celastrol for degradation in non-small-cell lung cancer. Carcinogenesis 2014;35:905-14.
- 82. Wang H, Teriete P, Hu A, Reveendra-Panickar D, Pendelton K, Lazo JS, *et al.* Direct inhibition of c-Myc-Max heterodimers by celastrol and celastrol-inspired triterpenoids. Oncotarget 2015;6:32380-95.
- 83. Mayola E, Gallerne C, Esposti DD, Martel C, Pervaiz S, Larue L, *et al.* Withaferin A induces apoptosis in human melanoma cells through generation of reactive oxygen species and down-regulation of Bcl-2. Apoptosis 2011;16:1014-27.
- 84. Yin R, Li T, Tian JX, Pan X, Liu RH. Ursolic acid, a potential anticancer compound for breast cancer therapy. Crit Rev Food Sci Nutr 2018;58:568-74.
- 85. Li Y, Zhang Y, Liu X, Wang M, Wang P, Yang J, *et al.* Lutein inhibits proliferation, invasion and migration of hypoxic breast cancer cells via downregulation of HES1. Int J Oncol 2018;52:2119-29.
- Qi WJ, Sheng WS, Peng C, Xiaodong M, Yao TZ. Investigating into anti-cancer potential of lycopene: Molecular targets. Biomed Pharmacother 2021;138:111546.
- 87. Chen QH, Wu BK, Pan D, Sang LX, Chang B. Beta-carotene and its protective effect on gastric cancer. World J Clin Cases 2021;9:6591-607.

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