Balanites aegyptiaca (L.) Del. (Hingot): A review of its traditional uses, phytochemistry and pharmacological properties

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Balanites aegyptiaca is an evergreen, woody, true xerophytic tree of tremendous medicinal importance. It belongs to the family Balanitaceae and is distributed throughout the drier parts of India. *B. aegyptiaca* has been used in a variety of folk medicines in India and Asia. Various parts of the plant are used in Ayurvedic and other folk medicines for the treatment of different ailments such as syphilis, jaundice, liver and spleen problems, epilepsy, yellow fever and the plant also has insecticidal, antihelminthic, antifeedant, molluscicidal and contraceptive activities. Research has been carried out using different in vitro and in vivo techniques of biological evaluation to support most of these claims. This review presents the traditional uses, phytochemistry and pharmacological properties of this medicinal plant.

Key words: Antibacterial, Balanites aegyptiaca, desert date, pharmacological properties, saponins, traditional uses

INTRODUCTION

Balanites aegyptiaca (L.) Del. belongs to the family Balanitaceae. It is a multibranched, evergreen tree distributed throughout the drier parts of India.^[1] It is widely grown in the Sudano-Sahielian region of Africa, the Middle East and South Asia.^[2] It is known by various names, e.g. Arabic names: Heglig (tree), lalob (fruit); trade name: zaccone, zachun, desert date (dried fruit); in India: Hindi name is Hingot and English name is thorn tree/desert date.^[4]

B. aegyptiaca is a mutibranched, spiny shrub or tree which grows up to 10 m in height. The leaves are alternate, two foliate, petioles are 3–6 mm long, leaflets are elliptic and have broadly pointed petioles up to 5 mm long. The spines of the plant are simple, straight, stout, rigid, green, alternate, supraaxillary, up to 5 cm long. Inflorescence is supraaxillary clusters or rarely supracemose. The flowers are small, bisexual, greenish white, fragrant, in axillary clusters, few or many in number, cymes or fascicles. The sepals are five in number (free), ovate and 3 mm long. The petals are five in number (two free), oblong-obovate, longer than the sepals. The stamens are ten in number, filaments glabrous, and anthers are dorsifixed. The ovary is ovoid, silky, five-celled and ovules are solitary in each cell, the style is short and conical. Fruit is an ovoid drupe, 2–5.6 cm long, found on a short thick stalk, and is faintly five grooved. The ripe fruit is brown or pale brown with a brittle coat enclosing a brown or brown-green

sticky pulp and a hard stone seed. Seeds are found exalbuminous and embryo is with thick plano-convex, or two-lobed cotyledons and a superior radical.^[4]

Traditionally, various parts of *B. aegyptiaca* have been reported to possess medicinal properties in different ethnobotanical surveys.^[1,5-8] It finds its place in the Ayurvedic pharmacopoeia of India and has also been described in some monographs, but none have described the complete traditional uses, phytochemistry and pharmacology of this plant. Therefore, we have compiled an up-to-date and comprehensive review of *B. aegyptiaca* that covers its traditional uses, phytochemistry and pharmacological properties.

TRADITIONAL USES

Various parts of *B. aegyptiaca* have their own traditional medicinal properties. This plant has been reported to be an antihelminthic, a purgative, vermifuge, febrifuge, emetic and can also cure other types of ailments like skin boils, leucoderma, malaria, wounds, colds, syphilis, liver and spleen disorders, and aches. [6] The bark of the plant is useful in curing mental diseases, epilepsy, yellow fever, jaundice and syphilis and can also act as a fumigant to heal circumcision wounds. [6] The boiled root of the plant can be used as a soup against stomach pain, anthrax, and the infusion of root also acts as an antidote to snake bite. [5] The infusion of root bark has been used in diarrhoea, in haemorrhoid and also acts as a fish poison. [8] The paste of shoot has been used for

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dressing of wounds and as tooth brushes when frayed. The thorns are used in the treatment of leprosy. Plant leaves are used in curing anthrax, for their antihelminthic activities and to clean malignant wounds. [6] The fruit can cure mouth ulcer, whooping cough, sleeping sickness and skin diseases. Fruit kernel has been found as a mild laxative, an antidote to arrow poison, and also acts as a vermifuge. [5] Kernel oil helps in curing skin disease. [11] The seeds are useful as ointments, to cure cough, colic pain and also have magicoreligious properties. [5,8]

PHYTOCHEMISTRY

The phytochemistry of its root, stem bark, leaves, fruit pulp, seed kernel, and mesocarp has been studied by different workers. *B. aegyptiaca* is a rich source of saponins. Saponins are glycosides consisting of sugar residues (one or more units of glucose, galactose, etc.) linked through oxygen with complex multiring compounds usually containing 27–31 carbon atoms. The aglycone part, which is also called sapogenin, is either a steroid (C27) or a triterpene

(C30). [9] Saponin containing plants are used in folk medicine, especially in Asia, and are intensively used in food, veterinary and medical industries. *B. aegyptiaca* contains different types of saponins, namely, balanitin -1, 2, 3, 4, 5, 6 and 7 [Figure 1]. The phytochemical description of the different parts is given below.

Root

Balanitin 1, 2 and 3, alkaloids and diosgenin have been isolated from the East African specimen.^[10-12] Diosgenin is a steroidal sapogenin (5-spirostan-3-ol) compound which is very useful in pharmaceutical industries as a natural source of steroidal hormones.^[13] Balanites roxburghii is an alternative source of diosgenin.^[14] The Indian species (*B. roxburghii*) contains the lowest level of both diosgenin and oil.^[15-17] The root wood of the plant contains Balanitism H.^[14]

Stem Bark

Balanitol (a new sesquiterpene) and the saponins, deltonin and protodeltonin, have been isolated from the bark of the Indian species (*B. roxburghii*).^[18] Furanocoumarin, bergapten

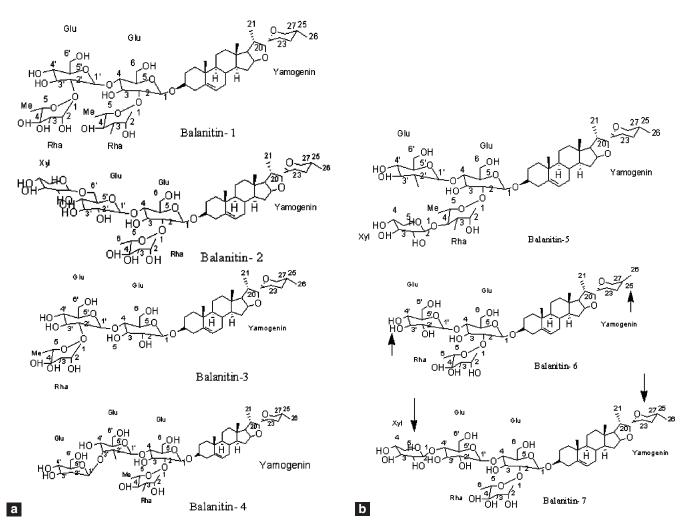


Figure 1: (a) Chemical structure of balanitin 1, 2, 3, and 4 isolated from different parts of *B. aegytptiaca*; (b) Chemical structure of balanitin 5, 6 and 7 isolated from different parts of *B. aegytptiaca*

and a dihydrofuranocoumarin (marmesin) have been isolated from the chloroform extract of the stem bark. [19] Balanitin 1, 2 and 3 have been isolated from East African species of *B. aegyptiaca* while diosgenin and sugars (glucose and rhamnose in the ratio 3:1) have been isolated from the Indian species (*B. roxburghii*). [10,20] Dicholoromethane extract has yielded two types of alkaloids N-*trans*-feruloyltyramine and N-*cis*-feruloyltyramine [Figure 2] and other metabolites like vanillic acid, syringic acid and 3 hydroxy-1-(4-hydroxy-3 methoxyphenyl)-1-propanone. [21]

Stem Wood

Balanitism 1 was isolated from the stem wood of the Indian species of *Balanites* (*B. roxburghii*).^[14]

Leaf

Six flavonides, glycosides identified as quercetin 3-glucosides, quercetin 3-rutinoside, 3-glucoside, 3-glucoside, 3-rutinoside, 3-7 diglucoside and 3-rhamnogalactosides of isorhamnetin have been extracted and identified from the leaves and branches of the Egyptian plant species.^[22]

Fruit

The fruit of *B. aegyptaica* consists of an epicarp, a mesocarp, an endocarp and a kernel.[23] The total saponin content has been found to be 7.2% in the mesocarp and 6.7% in the kernel. [24] Balanitin A, B, C, D, E and Balanitin F and G have been isolated from pulp and kernel, respectively. [14,25] The oil extracted from the kernel constituted 44-51% w/w and is composed of mainly triglycerides, with small quantities of diglycerides, phytosterols, sterol esters and tocopherols. [26] Besides, a known spirostanol glycoside, balanitin-3, and a new sapogenol, 6-methyldiosgenin, a new furostanol saponin, balanitoside and two pregnane glycosides have been isolated from the fruits (mesocarp) of B. aegyptiaca.[27-29] Spectroscopic and chemical analysis suggested the structure of the glycoside as 26-O-β-Dglucopyranosyl-3β,22,26-trihydroxy-furost-ene, 3-O-α-Lrhamnopyranosyl- $(1\rightarrow 2)$ -β-D-glucopyranosyl- $(1\rightarrow 4)$ -β-Dglucopyranoside and the saponins present in the mesocarp of B. aegyptiaca fruit are a mixture of 22R and 22S epimers of 26-(O-β-D-glucopyranosyl)-3-β-[4-O-(β-D-glucopyranosyl)-2-O-(α-L-rhamnopyranosyl)-β-D-glucopyranosyloxy]-22,26dihydroxyfurost-5-ene.[30]

Seeds

The seeds of *B. aegyptiaca* yielded four new cytostatic saponins, namely, balanitins 4, 5, 6 and 7.^[31] Seeds of *B. aegyptiaca* also contain deltonin and isodeltonin (steroidal spirostanol glycosides) which are used as molluscicidal agents.^[32]

PHARMACOLOGICAL PROPERTIES

The pharmacological activity, parts used, constituent

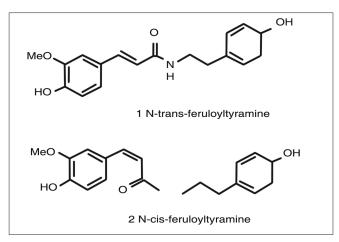


Figure 2: Structure of two alkaloids isolated from stem bark

compound responsible for pharmacological properties and sources of information are shown in Figure 3.

Insecticidal Activity

B. aegyptiaca acts as a potential natural larvicidal agent against mosquito larvae due to the larvicidal activities present in the saponin rich extracts in the various tissues such as fruit pulp, kernel, root, bark and leaf.[33-37] The water extracts of fruit kernel of B. aegyptiaca were found to be effective against the larvae of Aedes arabiensis, Culex quinquefasciatus and Aedes aegypti and it was concluded that A. arabiensis larvae were the most susceptible, followed by C. quinquefasciatus and then A. aegypti.[33] The root extract of the plant was found to be most lethal, followed by the bark among the various parts tested (fruit pulp, seed kernel, root, bark and leaves).[35,36] On the basis the very low adult emergence (only 18%) on using only 50 ppm of the root derived callus, saponins of B. aegyptiaca showed a great possibility for drastically reducing the A. aegypti population in the concerned areas.^[38] The main reason behind the larvicidal activity of plant extract may be the interaction of saponin molecules with cuticle membrane of the larvae, ultimately disarranging these.[39]

Antimicrobial Activity

Different studies were carried out to determine the antibacterial and antifungal activities of whole plant, root bark, stem bark, fruit mesocarp and leaves of *B. aegyptiaca*. [40-46]

Antibacterial activity

The leaf extracts of *B. aegyptiaca*, prepared in aqueous and organic solvents (acetone and ethanol), were tested for their antibacterial activity against *Salmonella typhi*, by using the disc diffusion method. Ethanolic extracts demonstrated higher antibacterial activity (16 mm zone of inhibition) while the aqueous extracts showed the least activity (4 mm zone of inhibition) at 100 mg/ml. The preliminary

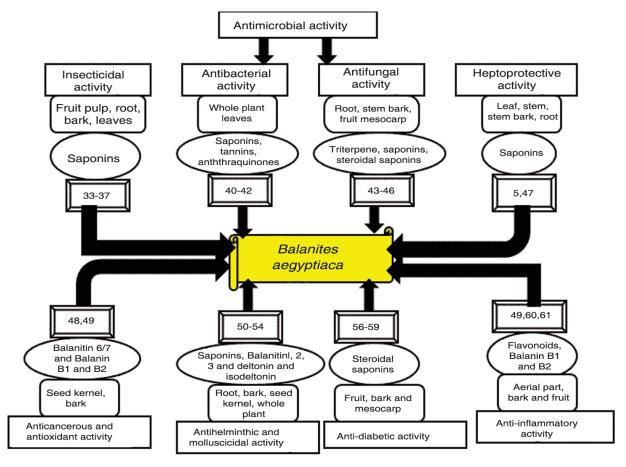


Figure 3: The pharmacological activity, parts used and constituents with sources of information

phytochemical analysis revealed the presence of saponins, tannins, phenols and anthraquinones in the extracts, and these were considered for antibacterial activity.^[40]

Methanolic and aqueous extracts of whole plant extract showed 4 mm inhibition zone in *Staphylococcus aureus* and 11 mm zone of inhibition in case of *Staphylococcus epidermidis*. ^[41] The extract of *B. aegyptiaca* supplemented with a 60–100 mg mineral (*kadosero*) revealed 100% reduction in bacterial colony in untreated well water. Chemical analysis of *kadosero* revealed the presence of $SO_4(0.0038 \text{ mg/g})$, $Fe^2(0.0027 \text{ mg/g})$, $Cl^-(232.683 \text{ mg/g})$ and $Na^+(151.25 \text{ mg/g})$. ^[42]

Antifungal Activity

Aqueous and methanolic (80%) extracts of root bark were screened for anticandidal activity by bioautography agar overlay method, using a standard strain of *Candida albicans* (ATCC 90028). These extracts revealed strong anticandidal activity. The identification of compounds responsible for the activity was not done. [43] The stem bark extracts isolated in various solvents were screened for their antifungal effects against *Aspergillus niger* and *C. albicans*, and these extracts also showed high antifungal activity against *C. albicans* (MFC 250 μg/ml). [44] The fruit mesocarp saponin

rich extract has been tested against common phytopathogenic fungi (*Pythium ultimum*, *Fusarium oxysporum*, *Alternaria solani*, *Colletotrichum coccodes* and *Verticillium dahliae*). The inhibitory effects of these extracts were measured *in vitro* and the concentrations that reduced the colony diameter of fungus to 50% of the control were determined. At 4% concentration, growth inhibitions were reported against *P. ultimum* (81.1%) and *A. solani* (34.7%).^[45] The antifungal activity may be due to presence of several triterpene saponins and steroidal saponins in *B. aegyptiaca*.^[46]

Hepatoprotective Activity

The extracts of leaf, stem, stem bark and root of *B. aegyptiaca* were screened for hepatoprotective activity in Wistar albino rats. The stem bark extracts of the plant showed significant (P < 0.05) hepatoprotective effects as revealed by a decrease in the activity of serum transaminase and alkaline phosphatase enzymes as compared to control rats.^[5] The effect of lyophilised extracts of *B. aegyptiaca* (1 g/kg) and silymarin (0.1 g/kg), a standard heptoprotective agent, given for 5 consecutive days, was tested on liver damage induced by paracetamol (0.6 g/kg) in the mice. *B. aegyptiaca* had a relatively modest hepatoprotective activity (27%) while silymarin protected about 92% of the

treated mice. [47] These results suggest that the extract could protect the paracetamol-induced liver damages perhaps by eliminating the deleterious effects of toxic metabolites from the drug.

Anticancerous and Antioxidant Activity

The mixture of balanitin-6 (28%) and balanitin-7 (72%) was evaluated in vitro for anticancer activity against six different human cancer cell lines, using the [3-(4, 5-dimethylthiazol-2yl)-diphenyltetrazolium bromide] colorimetric assay and in vivo in the murine L1210 leukaemia model. The mixture has demonstrated appreciable anticancer affects in human cancer cell lines in vitro as it displayed higher antiproliferative activity than etoposide and oxaliplatin but markedly lesser activity than taxol. The in vitro anticancer activities result at least partially from depletion of ATP, leading in turn to major disorganisation of actin cytoskeleton, ultimately resulting in the impairment of cancer cell proliferation and migration. In vivo, bal6/7 increased the survival time of mice bearing murine L1210 leukaemia grafts to the same extent as that reported for vincristine. These preliminary in vivo data suggest that it may be possible to generate novel hemi-synthetic derivatives of balanitin-6 and -7 with potentially improved in vitro and in vivo anticancer activity and reduced in vivo toxicity, thus markedly improving the therapeutic ratio.[48]

Balanitin B1 and B2, isolated from methanol and butanol extracts of *B. aegyptiaca* bark, have been evaluated *in vitro* and *in vivo* using a method based on the Briggs-Rauscher (BR) oscillating reaction and this revealed the antioxidant activity.^[49]

Antihelminthic and Molluscicidal Activity

The root, bark, seed kernel, fruit and whole plant extracts were found to be lethal to snails, miracidia and cereariac of schistosomes in various studies.^[50,51] A mixture of deltonin and 25-isodeltonin extracted from seeds was found to be molluscicidal against snail species Biomphalaria glabrata. [52] The antihelminthic properties of extract of *B. aegyptiaca* were compared with those of albendazole and praziquantel. [53,54] The efficacy of therapeutics of aqueous mesocarp extract against Fasciola gigantica in goat was 93.20-97.7%. The characteristic lesions of liver fasciolosis, egg count per gram of faeces (EPG), packed cell volume (PCV) and total blood count were significantly different from those of control and treated groups (P < 0.05).^[53] The efficacy of B. aegyptiaca fruit mesocarp (200 mg/kg) was compared with that of praziquantel (200 mg/kg) in mice infected with Sudanese strain of Schistosoma mansoni. A significant reduction was observed in EPG, egg burden in tissues and recovery of adult worms (P < 0.05) for both the extract- and the drugtreated animals.[54]

Antiparasitic

The crude methanolic extract has been found to have a moderate biological activity on *Leishmania major* in an *in vitro* study.^[55]

Antidiabetic Activity

The bark extract of B. aegyptiaca has been also shown to have a moderate effect on the activity of α -amylase which is responsible for the degradation of oligosaccharides.^[56] B. aegyptiaca fruit extracts (1.5 g/kg bw) reduced the blood glucose level by 24% and significantly decreased the liver glucose-6-phosphatase activity in diabetic rats.[57] The water and ethanolic extracts of B. aegyptiaca fruit extract induced significant reduction in serum glucose, glucagon, total lipids, total cholesterol, triglycerides level and transaminases [aspartate aminotransferase (AST), alanine aminotransferase (ALT) and yGT (gamma aminotransferase)] activities.[58] An aqueous extract of mesocarps of the fruits of B. aegyptiaca exhibited a prominent antidiabetic activity on oral administration in streptozotocin-induced diabetic mice. It is believed that the antidiabetic activity was due to the presence of steroidal saponins in the extracts.^[59]

Anti-inflammatory Activity

The ethanolic extract of aerial parts of *B. aegyptiaca*, when given orally as a suspension at 300 mg/kg bw per day, reduced the paw volume by 55.03%, whereas in the case of administration of 600 mg/kg bw per day it was 65.54%, indicating that the effect was dose dependant. The significant anti-inflammatory activity was evaluated in methanolic and ethanolic extracts of the bark in two different animal models, the carrageenan-induced oedema in the rat, and acetic acid-induced writhing test in mice. [49] Ethanolic extract of fruit of *B. aegyptiaca* also exhibited a proinflammatory activity. [60] The phytochemicals responsible for these activities were found to be flavonoids, saponins B1 and B2 isolated from bark and aerial parts of the plant. [49,61]

CONCLUSION

B. aegyptiaca is an evergreen tree distributed throughout the drier parts of India. It has been used since ages by the tribes of Rajasthan and Haryana to cure several diseases like whooping cough, sleeping sickness, guinea worm diseases and skin disorders. Economically also, it is a very significant plant because various parts of this plant contain saponins, alkaloids and diosgenin (secondary metabolites that have high pharmacological importance). The pharmacological studies of B. aegyptiaca demonstrated insecticidal, antibacterial, antifungal, hepatoprotective, anticancerous, antihelminthic, antiparasitic, antidiabetic and anti-inflammatory activities of the plant. Research carried out using different in vitro and in vivo techniques of biological evaluation support most of these claims. B. aegyptiaca may also act as a potential natural larvicidal agent

against mosquito larvae, owing to the presence of saponins which are effective in mosquito control, safe to mammals and available in high concentration in *B. aegyptaica*. So, this plant can be used as a drug in mosquito control.

India can benefit enormously if we can build a Golden Triangle among Modern Science, Modern Medicine and Traditional Medicine. [62] Indeed, triangles are a popular concept in complementary medicine, [63] but for AYUSH, the Golden Triangle presents a golden opportunity to bring these different systems together. Numerous drugs have entered the international market through exploration of ethnopharmacological and traditional medicine. Although scientist studies have been carried out by the scientists on many of the Indian botanicals, considerably small number of marketable drugs or phytochemical entities has entered the evidence-based therapeutics. E fforts are therefore needed to establish and validate safety and practice of Ayurvedic medicines. [64]

REFERENCES

- Anon. The useful plants of India. New Delhi: Publications and Information Council of Scientific and Industrial Research; 1986.
- Hall JB, Walker DH. Balanites aegyptiaca Del. a monograph. School of Agriculture and Forest Sciences. Banger: University of Wales; 1991
- 3. Hardman R, Sofowora EA. Balanites aegyptiaca as a source of steroidal sapogenins. Econ Bot 1972;26:169-73.
- Kirtikar KR, Basu BD. Indian medicinal plants. Vol. 1. Allahabad: Lalit Mohan Basu Publications; 1933.
- Ojo OO, Nadro MS, Tella IO. Protection of rats by extracts of some common Nigerian trees against acetaminophen-induced hepatotoxicity. Afr J Biotechnol 2006;5:755-60.
- Hamid O, Wahab M, Hassan E. Balanites aegyptiaca extract for treatment of HIV/ AIDS and leukemia. International Publication Number WO 2001/49306 A1.
- Neuwinger HD. Plants used for poison fishing in tropical Africa. Toxicon 2004;44:417-30.
- Bukar A, Danfillo IS, Adeleke OA, Ogunbodede EO. Traditional oral health practices among Kanuri women of Brono state Nigeria. Odontostomatol Trop 2004;27:25-31.
- Hostettmann K, Marston A. Saponins (chemistry and pharmacology of natural's products). Cambridge: University press; 1995.
- Liu HW, Nakanishi K. The structures of balanitins, potent molluscicides isolated from Balanites aegyptiaca. Tetrahedron 1982;38:513-9.
- 11. Keir YM. Investigation of certain plants used in Sudanese folk-medicine. J Afr Med Plants 1987;6:79-105.
- Gaur VS, Emmanuel CJ, Kant T. Direct *In vitro* shoot morphogenesis in desert date- B. aegyptaica (L.) Del. from root segments multipurpose trees in the tropics: Management and improvement strategies. In: Tewari VP, Srivastava RL, editors. Jodhpur, Scientific Publication. 2005. p. 701-4.
- 13. Liu MJ, Wang Z, Ju Y, Wong RN, Wu QY. Diosgenin induces cell cycle arrest and apoptosis in human leukemia K562 cells with the disruption of Ca2+ homeostasis. Cancer Chemother Pharmacol 2005;55:79-90.
- 14. Varshney IP, Vyas P. Saponin and sapogenin contents of Balanites aegyptiaca. Int J Crude Drug Res 1982;10:3-7.

- Chapagain BP, Wiesman Z. Determination of saponins in the kernel cake of Balanites aegyptiaca by HPLC-ESI/MS. Phytochem Anal 2007;18:354-62.
- Chapagain BP, Wiesman Z. Variation in diosgenin level in seed kernels among different provenances of Balanites aegyptiaca Del (Zygophyllaceae) and its correlation with oil contents. Afr J Biotechnol 2005;4:1209-13.
- 17. Desai KG, Desmukh UK, Saoji AN. Studies on diosgenin contents of B.aegyptaica. East Pharm 1978;21:191-3.
- Cordano G, Terrien MA, Plonsky J, Rabanal RM, Varenne P. Balanitol, a new sesquiterpene from B.roxburghii, carbon-13 NMR analysis of eudesonance sesquiterpenoids. J Indian Chem Soc 1978;55:1148-51.
- Ahmed AS, Kingham AD, Geoffery AC, Norman RF. Isolation of bergapten and marmesin from B. agyptica. Planta Med 1981;43: 92-103
- Jain DC. Antifeedant active saponins from Balanites roxburghii stem bark. Int J Crude Drug Res 1978;6:45-7.
- 21. Sarker SD, Bartholomew B, Nash RJ. Alkaloids from Balanites aegyptiaca. Fitoterapia 2000;71:328-30.
- 22. Maksoud SA, El-Hadidi MN. The flavonoids of Balanites aegyptiaca from Egypt. Plant Syst Evol 1988;160:153-8.
- Mohamed AM, Wolf W, Spieb WE. Physical, morphological and chemical characteristics, oil recovery and fatty acid composition of Balanites aegyptiaca Del. Kernels. Plant Foods Hum Nutr 2002;57:179-89.
- 24. Watt JM, Brandwijk BM. The medicinal and poisonous plants of southern and eastern Africa. London: Livingstone Ltd.; 1962.
- Varshncy IP, Janin DC. Study of glycosides from T. foenumgraccum L. leaves. Nad Acad Sci Lett (India) 1979;2:331-2.
- Abu-El-Futuh IM. Balanites aegyptiaca, an unutilized raw material potentially ready for agro-industrial exploitation. United nations Industrial Development Organization, Vienna, Austria report UNIDO, 1983. p. 10-494.
- 27. Hosny M, Khalifa T, Caliş I, Wright AD, Sticher O. Balanitoside, a furostanol glycoside, and 6-methyldiosgenin from Balanites aegyptiaca. Phytochemistry 1992;31:3565-9.
- Kamel MS. A furostanol saponin from fruits of Balanites aegyptiaca. Phytochemistry 1998;48:755-7.
- Kamel MS, Koskinen A. Pregnane glycosides from fruits of Balanites aegyptiaca. Phytochemistry 1995;40:1773-5.
- Staerk D, Chapagain BP, Lindin T, Wiesman Z, Jaroszewski JW.
 Structural analysis of complex saponins of Balanites aegyptiaca by 800 MHz 1H NMR spectroscopy. Magn Reson Chem 2006;44:923-8.
- 31. Pettit GR, Doubek DL, Herald DL, Numata A, Takahasi C, Fujiki R, *et al.* Isolation and structure of cytostatic steroidal saponins from the African medicinal plant Balanites aegyptiaca. J Nat Prod 1991;54:1491-502.
- 32. Gnoula C, Guisso P, Duez P, Frederich M, Dubois J. Nematocidal compounds from seeds of Balanites aegyptiaca Isolation and structure Elucidation. Int J Pharmacol 2007;3:280-4.
- Zarroug IM, Nugud AD, Basir AK, Mageed AA. Balanites aegyptiaca as a mosquito larvicides. Int J Crude Drug Res 1990;28:267-71.
- 34. Wiesman Z, Chapagain BP. Laboratory evaluation of natural saponins as bioactive agents against Aedes aegypti and Culex pipiens. Dengue Bull 2003;27:168-73.
- Chapagain B, Wiesman Z. Larvicidal effects of aqueous extracts of Balanites aegyptiaca (desert date) against the larvae of Culex pipiens mosquitoes. Afr J Biotechnol 2005;4:1351-4.
- 36. Wiesman Z, Chapagain BP. Larvicidal activity of saponin containing extracts and fractions of fruit mesocarp of Balanites aegyptiaca. Fitoterapia 2006;77:420-4.
- 37. Weisman Z. Balanites aegyptica saponins and uses thereof.

- International Publication Number WO 2006/137069 A2.
- Chapagain BP, Saharan V, Wiesman Z. Larvicidal activity of saponins from Balanites aegyptiaca callus against Aedes aegypti mosquito. Bioresour Technol 2008;99:1165-8.
- Morrissey JP, Osbourn AE. Fungal resistance to plant antibiotics as a mechanism of pathogenesis. Microbiol Mol Biol 1999;63:708-24.
- Doughari JH, Pukuma MS, De N. Antibacterial effects of Balanites aegyptiaca L. Drel. and Morianga oleifera Lam. On Salmonella typhi. Afr J Biotechnol 2007;6:2212-5.
- Parekh J, Chanda S. In vitro screening of antibacterial activity of aqueous and alcoholic extracts of various Indian plant species against selected pathogens from Enterobacteriaceae. Afr J Microbiol Res 2007;1:92-9.
- 42. Otieno JN, Hosea KM, Lyaruu HV. The effect of local minerals Kadsaro towards the antimicrobial activity of medicinal plants extract. Case of Lake Victoria Basen, Tarim Tanzania. Afr J Tradit Complement Altern Med 2007;4:1-6.
- Runyoro DK, Matee MI, Ngassapa OD, Joseph CC, Mbwambo ZH.
 Screening of Tanzanian medicinal plants for anti-Candida activity.
 BMC Complement Altern Med 2006;30:6-11.
- 44. Maregesi SM, Pieters L, Ngassapa OD, Apers S, Vingerhoets R, Cos P, et al. Screening of some Tanzanian medicinal plants from Bunda district for antibacterial, antifungal and antiviral activities. J Ethnopharmacol 2008;119:58-66.
- Chapagain BP, Wiesman Z, Tsror L. In vitro study of the antifungal activity of saponin-rich extracts against prevalent phytopathogenic fungi. Afr J Tradit Complement Altern Med 2007;4:1-6.
- Farid H, Haslinger E, Kunert O, Wegner C, Hamburger M. New steroidal glycosides from Balanites aegyptiaca. Helv Chim Acta 2002;85:1019-26.
- 47. Ali BH, Bashir AK, Rasheed RA. Effect of the traditional medicinal plants Rhazya stricta, Balanitis aegyptiaca and Haplophylum tuberculatum on paracetamol-induced hepatotoxicity in mice. Phytother Res 2001;15:598-603.
- 48. Gnoula C, Megalizzi V, Neve ND, Sauvage S, Ribaucour F, Guissou P, et al. Balanitin-6 and 7: Diosgenyl saponins isolated from Balanites aegyptiaca Del. display significant anti-tumor activity *in vitro* and in vivo. Int J Oncol 2008;32:5-15.
- Speroni E, Cervellati R, Innocenti G, Costa S, Guerra MC, Dall S, et al. Anti-inflammatory, anti-nociceptive and antioxidant activities of Balanites aegyptiaca (L.) Delile. J Ethnopharmacol 2005;98:117-25.
- Ndamba J, Robertson I, Lemmich E, Chandiwana SK, Furu P, Molgaard P. Investigation of the diurnal ontogenic and seasonal variation in the molluscicidal saponins content of Phytolacca dodecancra aqueous berry extracts. Phytochemistry 1994;35:95-9.

- 51. Guyatt HL, Evens D. Economic consideration for helminthes control. Parasitol Today 1992;8:397-402.
- 52. Brimer L, ElSheikh SH, Furu P. Preliminary investigation of the disposition of the molluscicidal saponin deltonin from Balanites aegyptiaca in a snail species (Biomphalaria glabrata) and in mice. J Pesticide Sci 2007;32:213-21.
- 53. Koko WS, Galal M, Khalid HS. Fasciolicidal efficacy of Albizia anthelmintica and Balanites aegyptiaca compared with albendazole. J Ethnopharmacol 2000;71:247-52.
- 54. Koko WS, Abdalla HS, Galal M, Khalid HS. Evaluation of oral therapy on Mansonial Schistosomiasis using single dose of Balanites aegyptiaca fruits and praziquantel. Fitoterapia 2005;76:30-4.
- 55. Fatima F, Khalid A, Nazar N, Abdalla M, Mohomed H, Toum AM, et al. In Vitro assessement of anti-cutaneous leismaniasis activity of some Sudaneses plants. Turkiye Parazitol Derg 2005;29:3-6.
- Funke I, Matthias F, Melzig CW. Phytotherapy in type 2 diabetes mellitus. Investigations of traditional herbal drugs as possible alpha amylase inhibitors. J Phytother Res 2005;26:271-4.
- 57. Mohamed ZG, Maha MS, Manal FI, Nibal DT. Biochemical study of the anti diabetic action of the Egyptian plants Fenugreek and Balanites. Mol Cell Biochem 2006;281:173-83.
- Zaahkouk SA, Rashid SZ, Mattar AF. Ant diabetic properties of water and ethanolic extracts of Balanites aegyptiaca fruits flesh in senile diabetic rats. Egyptian J Hosp Med 2003;10:90-108.
- Kamal MS, Ohtani K, Kurokawa T, Assaf MH, Shanawany MA, Ali AA, et al. Studies on B.aegyptaica fruits an ant diabetic Egyptian folk medicine. Chem Pharm Bull (Tokyo) 1991;39:1229-33.
- Koko WS, Mesaik MA, Yousaf S, Galal M, Choudhary KI. *In vitro* immunodulating properties of selected Sudanese medicinal plants. J Ethnopharmacol 2008;118:26-34.
- 61. Gaur K, Nema RK, Kori ML, Sharma CS, Singh V. Antiinflammatory activity of Balanites aegyptiaca in experimental animals models. Int J Green Pharm 2008;2:214-7.
- 62. Mashelkar RA. Second world Ayvrveda congress (Themes: Ayvrveda for the future) Inaugural address: Part I. Evid Based Complement Alternat Med 2008;5:129-31.
- Cooper EL. CAM, eCAM, Bioprospecting: The 21st Century Pyramid. Evid Based Complement Alternat Med 2005;2:125-7.
- Patwardhan B, Warude D, Pushpangadan P, Bhatt N. Ayurveda and traditional Chinese medicine: A comparative overview. Evid Based Complement Alternat Med 2005;2:465-73.

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