Antihyperlipidemic and cardioprotective effects of plant natural products: A review

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Abstract

The present review article explains antihyperlipidemic and cardioprotective effects of various plant natural products. These are antioxidants which found in vegetable oils, fruits, seeds, plant leaves, stem, and plant roots. This article emphasizes therapeutic potential of sulfur-containing compounds, anthraquinones, thiosulfinates and ajoenes, tricin, quercetin, Kaikasaponin III and tectorigenin, polyphenols, vitamins and minerals, flavonoid, and phenolics mainly naringenin; aglycone, flavanone (+)-catechin, (-)-epicatechin, and procyanidin B2, quercetin, kaempferol, myricetin, rutin, naringenin, catechin, fisetin, and gossypetin, thymol and carvacrol, ceramicine B (limonoid), ginsenoside Rb1 and Rb2, naphthalenic compounds 6-methoxysorigenin and its glycosides, acylates, madecassoside (terpenoids), Miroestrol (phytoestrogen), quercetin rutinoside (rutin), tournefolal, and tournefolic acids A and B. Few other compounds such as thiobarbituric acid, niacin, hederagenin monodesmosides, kalopanaxsaponin A (KPS-A) and sapindoside C, abietane and seco-abietane diterpenes, lycopene, Curcurbitacins A, B, C, D, E, I, J, K, and L, and Colocynthosides A and B were found active. For a healthy life regular intake of plant natural products or herbal diets significantly reduce the cardiovascular risks. Before use, well-designed clinical trials are essentially required to explore real efficacy and safety of these plant products. For management of cardiovascular risks lipid profile is highly needful at an earlier stage. In addition, microlevel biomolecular changes responsible for multiple morbidities affiliated to CVD diseases must be measured to find minor changes at an earlier stage.

Key words: Antilipidemic herbal preparations, cardiovascular diseases, hyperlipidemia, LDL cholesterol, plant natural products

INTRODUCTION

due to bad food habits hyperlipidemia, diabetic cardiomyopathy and cardiovascular diseases (CVDs) become serious global health problems. Cases related to obesity, hyperlipidemia, atherosclerosis, and diabetic cardiomyopathy (DCM) are increasing day by day and large numbers of deaths are occurring worldwide.[1] There are two important reasons of these CVD diseases and disorders; first one is metabolic storage of excess fat/lipids (adipogenesis) in body tissues after dietary use, second geneticor heredity-related disorders. Hyperlipidemia is the main cause of generation of CVDs myocardial infarction and strokes. It is a major pathological condition associated with disrupted lipid levels and physiological redox homeostasis. Increased cholesterol levels create problems in streaming of blood through blood vessels. This hypercholesterolemic state causes multiple risks to cardiovascular system impose excessive release of reactive oxygen species

(ROS) that lead to enhanced lipid peroxidation.^[2] It also aggravates atherosclerosis and oxidative stress in patients.^[2] Today, most of the people are affected due to obesity induced blood pressure, blood vessel inflammation arteriosclerosis, and diabetic cardiomyopathy (DCM). This is a more common and severe complication of diabetes that is causing high mortality in association of blood pressure.^[3]

Cardiovascular diseases (CVDs) and other associating disorders such as myocardial infraction, hypertension, peripheral vascular diseases, coronary heart disease, cardiomyopathies, and dyslipidemias become significant health burden and most frequent causes of morbidity and mortality in man [Figure 1].^[4] More especially in vascular

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Received: 10-01-2021 **Revised:** 03-03-2021 **Accepted:** 12-03-2021 diseases like atherosclerosis atherogenic plaques are formed as a result of accumulation of lipids and fibrous elements in the subendothelial space of large arteries and subsequent formation of lesions inside the coronary and cerebral arteries.^[5] Pathogenesis of atherosclerosis is multifactorial, and many modifiable and non-modifiable risk factors have been identified.^[6] Atherosclerosis is a chronic multifactorial disease characterized by mainly changes of blood lipids profile and inflammation in vessel wall. It is well known that lipid peroxide in low-density lipoprotein (LDL) plays an important role in atherosclerosis and atherosclerotic diseases. Therefore, timely prevention and therapy of atherosclerosis is highly essential to reduce the risk of the development of its clinical manifestations.^[7] These risk factors collectively contribute to the development, progression, and rupture of atherosclerotic plaque.[8]

Natural plant products can be used for the treatment or prevention of cardiovascular and metabolic disorders.[9] These plant-derived antioxidants^[7,10] can play an essential role in the prevention of cardiovascular diseases[11-13] mainly atherosclerosis.^[14] Due to its strong free radical scavenging activity, rejuvenation of health is possible if used in dietary supplements. Plant natural products become valuable source of less toxic, highly efficacious, and safe multitarget drugs. These are rich in antioxidants if used in dietary supplements. Plant flavonoids stop hardening and narrowing of the arteries in atherosclerosis and also act as anti-atherogenic, hypotensive, lipid-lowering, and anti-thrombotic agents. These subsequently reduce the chance of strokes, heart attacks, and peripheral vascular diseases [Figure 2]. Thus, herbs can potentially reduce serum lipid profile if include in dietary habits.[14]

Natural products promote health and successfully prevent cardiovascular and metabolic disease. No doubt these are valuable source for the discovery of new drugs [Figure 2]. However, there is a need to explore the cellular and molecular cardioprotective mechanisms of herbs used in treatment

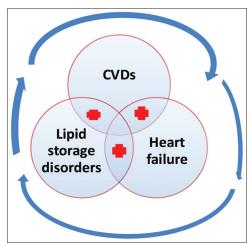


Figure 1: Inter-relationship between CVD, lipid storage disease, and heart failure

CVDs and their risk factors. However, before its use, lipid profile mainly serum concentrations of total cholesterol, LDL-C, HDL-C, triglycerides, apoA and apoB, SGOT, SGPT, glucose, and insulin must be checked. [14] It is advised that only pharmacologically approved herbs should be prescribed for therapeutic use. More often, herb-drug interactions should be investigated and checked thoroughly for their efficacy, safety, and toxicity. [15] This review emphasizes safe use of natural plant products and herbal medicines for the treatment of hypercholesterolemia and CVDs.

SOURCE OF INFORMATION

For writing this comprehensive research review on antihyperlipidemic and cardioprotective effects of various plant natural products, various databases were searched. For collection of relevant information, specific terms such as medical subject headings (MeSH) and key text words, such as "antihyperlipidemic and cardioprotective," "natural plant products," and therapeutic uses" published till 2020 were used in MEDLINE. Most specially for retrieving all articles pertaining to the natural plant products and herbal medicines for the treatment of hypercholesterolemia and CVDs, electronic bibliographic databases were searched and abstracts of published studies with relevant information on the venom toxins/allergens were collected. Furthermore, additional references were included through searching the references cited by the studies done on the present topic. Relevant terms were used individually and in combination to ensure an extensive literature search. For updating the information about a subject and incorporation of recent knowledge, relevant research articles, books, conferences proceedings, and public health organization survey reports were selected and collated based on the broader objective of the review. This was achieved by searching databases. including SCOPUS, Web of Science, and EMBASE, PubMed, PMC, Publon, Swiss-Prot, and Google searches. From this common methodology, discoveries and findings were identified and summarized in this final review.

USE OF VEGETABLE OILS

Vegetable oils are edible and typical dietary component and predominantly used for cooking purposes. These are extracted from some seeds, nuts, cereal grains, and fruits and remain in liquid state at ambient temperatures. [16] Vegetable oils are mainly triacylglycerols (92–98%), polar lipids (phospholipids and galactolipids), monoacylglycerols, and diacylglycerols. These also contain free fatty acids and polyisoprenoids in minor amounts. [17] Other minor components are tocopherols/tocotrienols (up to 900 mg kg⁻¹) and phytosterol esters/phytosterols (up to 1%). Plant oils contain wide variety of fatty acids which are nutritionally beneficial to human health. ALA (alpha-linolenic acid), another omega-3 fatty acid, is found in plant sources such as nuts and seeds mainly flaxseed.

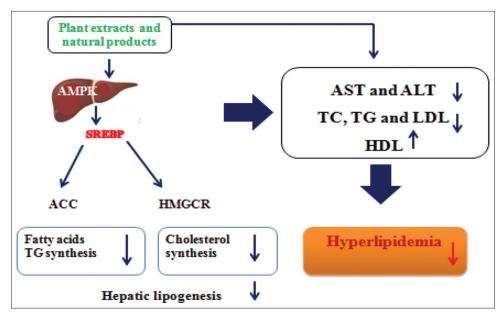


Figure 2: Possible mechanisms in hepatic tissues showing action of plant natural products that prevent hyperlipidemia through AMPK pathway and regulation of lipid metabolism. *AMPK – Adenosine monophosphate-activated protein kinase, ACC – Acetyl-CoA carboxylase, HMG-CoA reductase, AST – Aspartate transaminase, ALT – Alanine transaminase, CVD – Cardiovascular disease, LDL – Low-density lipoprotein cholesterol, TC – Total blood cholesterol (TC) and TG – Triglyceride

Vegetable oils contain non-volatile organic compounds (VOCs) which are less toxic and less dissolving power of non-polar and polar bioactive components with the function of fatty acids and/or lipid classes in vegetable oils, other minor components.^[18] Natural products can be used to have new drugs as they possess a variety of pharmacological targets. [19] Vegetable oils are source of unsaturated fatty acids which can replace deposited fat; vegetable oils also possess natural antioxidants which can extract out or sequestration of bad cholesterol and fats. Plant oils can easily leach out deposited fat from tissue adipocytes [Figure 2]. Peroxisome proliferator-activated receptor γ (PPARγ), the chief regulator of adipogenesis, has been acutely investigated as a molecular target for natural products in the development of antiobesity treatments. Due to their susceptibility to oxidation from the exposure to oxygen, heat and light, resulting in the formation of oxidation products, such as peroxides and hydroperoxides and plant oils rich in polyunsaturated fatty acids have a limited shelf life.[20,21]

Most vegetable oils are unstable during storage or cooking due to the oxidation of their polyunsaturated fatty acids. Therefore, partial hydrogenation is frequently applied to improve the flavor and oxidative stability of vegetable oils by reducing the content of the highly unsaturated linolenic and linoleic acids. [20,21] Vegetable oils contain predominantly unsaturated (light, liquid) fatty acids of two kinds: Monounsaturated (oleic acid – mainly in extra virgin olive oil) and polyunsaturated (linoleic acid and linolenic acid – in oils extracted from oil seeds). Plant origin short-chain omega-3 fatty acids are ALA (alpha-linolenic acid) mainly EPA eicosapentaenoic acid and DHA docosahexaenoic acid extend big health benefits. These are extracted

from flaxseed.^[20,21] Besides, plant oils "Fish oil" are also nutritionally and medically important as they contain long-chain polyunsaturated fatty acids. Omega-3 fatty acids are found in fatty layers of cold water fish and shellfish, plant, and nut oils. Fish are low in saturated fat. These provide benefits to cardiovascular disease patients [Figure 3].^[20,21]

USE OF NATURAL ANTIOXIDANTS

Plants contain natural antioxidants mainly flavonoids phenolic oligomeric proanthocyanidins (OPC) and pterostilbene (PT) with niacin (NA). These natural antioxidants blends show potential antihyperlipidemic effects and assist in reduction in lipid peroxidation, decrease cellular damage, and contribute cardioprotective activity.[22] This is the main reason that medicinal plants constitute a principal health-care resource. Due to their multitarget therapeutic action, there is a gradual increase in acceptance at the global level. Today, there are number of herbal medicines which are used for the treatment of hypercholesterolemia and cardiovascular problems mainly CVDs. There are confirmed scientific reports on usage of herbal medicines which prove their therapeutic efficacy. These simple natural antioxidants blends present in drug formulations can help in mitigating chronic illnesses in general populations.

CONSUMPTION OF VEGETABLES AND FRUITS

Vegetables and fruits are source of variety of polyphenols, vitamins, and minerals which act as natural antioxidants and

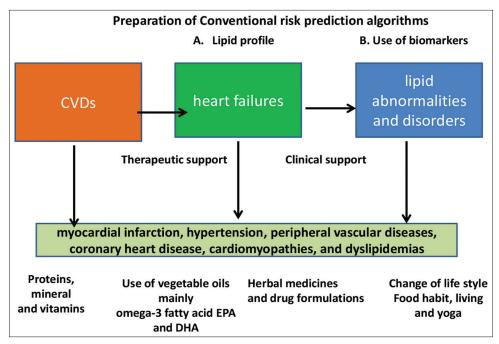


Figure 3: Management of hyperlipidemia mainly CVDs, heart failure, and lipid abnormalities using various natural plant products

assist in prevention of hyperhomocysteinemia. Drinking juices are a potential way to improve cardiovascular health.[23] Herbal tea (ZHT) for antihypercholesterolemic and antilipidemic perspectives in decrease of serum LDL cholesterol (5.6%) and TG (12.5%) was also observed after consumption of T3 at day 60.[24] Pistacia atlantica subsp. Kurdica shows antihyperlipidemic and antioxidative activity in streptozotocin-induced diabetic mice.[25] Blue-green algae (Nostoc commune) inhibit cholesterol absorption in mice and lower down plasma total cholesterol and triglyceride (TG) levels compared with controls.[26] Interestingly, kombucha inhibit absorption of LDL cholesterol and triglycerides and cause a significant increase in HDL cholesterol life.[27] Both zedoary (Curcuma zedoaria Roscoe.) and Xanthium strumarium L. (Asteraceae) plant extracts showed antilipidemic effects [Figure 3].[28]

USE OF HERBAL MEDICINES

The use of medicinal herbs continues to be an alternative treatment approach for several diseases including CVDs. At present, use of herbal preparations has been increased because of their therapeutic efficacy. Herbal preparations and drugs are cost effective and show big therapeutic promise compared to standard modern therapies. Due to their easy usage and less toxic, there is a general belief among people that they are safe. Polyherbal formulations prepared using different proportions of *Commiphora mukul* (Hook ex Stocks) Eng., *Salacia reticulata* Wight, *Terminalia arjuna* (Roxb.) Wight and Arn, and *Curcuma longa* Linn. showed antilipid-peroxidative effects. [29] Today, statins and biguanides are two prominent representatives of natural products which are mostly used against metabolic disease.

There is going on a large search for lipid-modifying agents. Few of them are statins, fibrates, niacin, and ezetimibe that significantly reduce the risk of atherosclerotic morbidities. These are established herbal drugs used in CVD treatments. There are important approaches such as inhibition of proprotein convertase, inhibition of the synthesis of apolipoprotein (apo) B, and microsomal triglyceride transfer protein to block the formation of atherogenic lipoproteins which occur are atherosclerosis. Other approaches are inhibition of adenosine triphosphate citrate lyase to inhibit the synthesis of cholesterol, inhibition of the synthesis of lipoprotein, apoC-III to reduce triglyceriderich lipoproteins, and cholesteryl ester transfer protein to enhance high-density lipoprotein (HDL) functionality.[30] Plant natural products inhibit cholesterol absorption in mice and lower down plasma total cholesterol and triglyceride (TG) levels [Figure 3].[31]

Various types of herbs can potentially reduce serum lipid levels through different metabolic pathways. More specially, the herb-drug interactions may decrease the morbidity if well-tested anti-hyperlipidemic drug formulations are being provided life.^[32] However, GSTC3 successfully decreased the serum total cholesterol, low-density lipoprotein (LDL), very low-density lipoprotein (VLDL), triglycerides, and phospholipids similar to standard atorvastatin while maintaining high-density lipoprotein (HDL) at normal levels at dose of 100 mg/kg body weight. Significantly lower values of LDL, VLDL, and atherogenic index of plasma as well as higher HMG-CoA/mevalonate ratios demonstrate significant hypolipidemic effect for GSTC3. It may be possibly due to partial inhibition of HMG-CoA reductase activity in experimental animals [Figures 2 and 3].^[32]

USE OF ANTILIPID PEROXIDATION NATURAL PLANT EXTRACTS

Different solvent extracts prepared from aerial parts of various plant species such as Bael leaf extracts, [33] Talinum triangulare (Jacq.) Willd., [34] Momordica charantia fruit extract Linn.,[35] and use of Aloe vera also decreases LDL cholesterol levels by 45%, 3%, and 69%, respectively, in experimental rats.^[36] Dietary use of onion (Allium cepa L.) significantly cut down the hyperlipidemia, hypertension, and diabetes.[37] Similar antilipid peroxidation activities are also reported in Padina boergesenii, [38] Maytenus royleanus, [39] Phyllanthus emblica leaves, [40] Tunisian Zizyphus lotus, [41] and Terminalia chebula Retz. (Combretaceae) fruits.[42] Hypericum x moserianum and Hypericum ericoides show potential antilipid peroxidation activities.[43] Methanolic extract of the bark of Ficus amplissima (FAB) shows significant antilipid-peroxidative effect in the pancreas of STZ-induced diabetic rats.^[44] Similar antilipid peroxidation activities also found in Sanguisorba officinalis methanolic, [45] and aqueous or ethanolic extracts, [46] Apocynum venetum (AVLE), [47] Piper trioicum Roxb. and Physalis minima L., [48] and Tunisian Rhamnus alaternus.[49] Various solvent extracts prepared from different plant species i.e. Phaseolus trilobus, [22] O. basilicum and T. foenum-graecum, [50] Asparagus racemosus (AR) and Premna integrifolia^[51] Linn, Murraya koenigii, [52] Gongronema latifolium, [53] (Utazi), and Jasminum grandiflorum Linn.^[54] Gongronema latifolium (Utazi), [54] Jasminum grandiflorum Linn. extracts. Extracts prepared from fruit pulp of the Chakapat lychee (Litchi chinensis Sonn.),[55,56] Rosa davurica Pall.,[57] Aegle marmelos (AMFEt), and Momordica charantia Linn. Fruits showed antilipid-peroxidative activity in streptozotocin diabetic rats.^[58] Hamsters fed on 3% or 9% tomato paste showed reduced rates of serum TC and LDL levels.[59]

Strong antilipid peroxidation activity is also reported in *Pinus* morrisonicola Hay. [60] and Selaginella involvens (Sw.) Spring, a wild fern water extract.^[61] The oral consumption of Gel-Et for 4 weeks caused significant reduction total cholesterol levels by 11%, 17%, and 25%, and LDL cholesterol levels by 69%. [62] Similarly, SCBe showed antilipidemic activity as evidenced by significant decrease in serum TC, TG, and LDL-C levels and significant increase in HDL-C level in antilipidemic activities of S. cochinchinensis bark methanolic extract (SCBe) in streptozotocin (STZ)-induced diabetic rats life. [63] Hypolipidemic activities are reported in Averrhoa bilimbi Linn. leaves (Oxalidaceae, Common name: Bilimbi), [64] Phyllanthus urinaria, [65] radix Angelica sinensis, [66] Yam (Dioscorea batatas) tuber mucilage, [67] Nardostachys jatamansi, [68] and Cortex Phellodendri. [69] Radix Aconiti Preparata and Radix glycyrrhizae can significantly suppress lipid peroxidation of myocardial homogenate^[70] and Ficus carica leaves also showed antilipidemic effects [Figure 3].^[71]

ANTIHYPERLIPIDEMIC EFFECTS

Garlic (Allium Sativum)

Garlic is widely used in Asian countries for dietary purposes. Both aged and processed garlic are used for the treatment of cardiovascular and diabetic patient's life. [72,73] It contains fructooligosaccharides which have potential benefits on health life.^[74] Homemade green garlic preparations possess cardioprotective potential to prevent cardiovascular diseases. This is the main reason that Mediterranean people who have a noted lower mortality rate due to cardiovascular diseases life. [75] Garlic shows multiple protective effects and improves functioning of cardiovascular system.[76-78] It removes off atherosclerosis and does reduction of serum lipids.[78] It shows inhibition of platelet aggregation and enhancement of fibrinolysis. Wild garlic (Allium ursinum) has been reported to contain similar amounts of sulfur-containing compounds, [79] thiosulfinates and ajoenes from garlic, exert similar effects on cyclooxygenase, 5-lipoxygenase, angiotensin-converting enzyme, and platelet aggregation.[34] DADS show strong antilipidemic effects [Table 1].[80]

Garlic products showed positive effects on lipid metabolism in cholesterol-fed rats^[81] and cut down lipid contents in experimental animals.^[82-84]. Possibly, it may occur through inhibition of 3-hydroxy-3-methyl-glutaryl-CoA reductase or other enzymes.^[85-87] More specifically, garlic-derived organosulfur diallyl di- and trisulfide compounds inhibit cholesterol biosynthesis in primary rat hepatocyte cultures.^[88-89] Moreover, garlic ingredients increase loss of bile salts in feces and mobilization of tissue lipids into circulation. Garlic does lowering of blood lipids, blood sugar, and fibrinogen and induces fibrinolytic activity in patients with coronary artery diseases. Garlic essential oil shows profound effect on postprandial hyperlipidemia and does prevention of atherosclerosis [Figures 2 and 3].^[90]

Wild garlic (A. ursinum) causes decrease hepatocyte cholesterol synthesis in vitro.[91] Aged garlic extract and its constituent SAC have been found to protect vascular endothelial cells from injury caused by oxidized LDL.[92] More often, aged garlic extract, "Kyolic," and essential oil[93] showed anti-atherosclerosis-related effects. [94] Garlic products affect lipid content in normal and atherosclerotic human aortic cells.[95] Garlic products show long-term effect on the development of plaque formation in the carotid branches of both femoral arteries[96] and lower down blood lipid level in cholesterol-fed rabbits.^[97] Daily supplementation of aged garlic extracts lowers down lipoprotein oxidation susceptibility^[98] and protects against LDL malfunction.^[99,100] Garlic contains oil-soluble organosulfur compounds which affect on doxorubicin-induced lipid peroxidation.[101] Garlic extracts cut down vascular tissue lipids, fatty streak formation, and atherosclerotic plaque size.[102] Garlic extract significantly normalizes the ratio of serum LDL to HDL

Upadhyay: Effects of plant natural products

Table 1: Important bio-organic ingredients isolated from various plant species which show antihyperlipidemic and cardioprotective activity

Plant natural product	Plant species	Plant part	Biological effects
Sulfur-containing compounds	Allium sativum,	Bulbs	Hypolipidemic and
	garlic		cardioprotective
Thiosulfinates and ajoenes	Oil-macerated garlic extract	Bulbs	Antihyperlipidemic
$Umbelliferon-\alpha\text{-}D\text{-}glucopyranosyl\text{-}(2I \rightarrow 1II)-\alpha\text{-}Dglucopyranoside}$	Aegle marmelos	leaf	Antihyperlipidemic
Anthraquinones	<i>Radix Rhizoma</i> Rhei	Leaf and buds	Antihyperlipidemic
Methyl ferulate, methyl p-hydroxycinnamate, p-hydroxybenzaldehyde, tricin, and quercetin 3,4'-dimethyl ether	Sorghum bicolor.	Stem	Antihypercholesterolemia and protective to CVD
Kaikasaponin III and tectorigenin	Pueraria thunbergiana	Flowers	Antilipid-peroxidative effects
N-acetylcysteine	Allium species	Bulbs	Antihypercholesterolemic
Gallic acid (trihydroxybenzoic acid	Gallnuts, sumac, witch hazel, tea leaves, oak bark	Stem bark	Antihypercholesterolemic
Polyphenols, vitamins, and minerals	Leafy green vegetables	Vegetables and fruits	Hyperlipidemic and cardioprotective
Oligostilbenes	Vitis amurensis	Leaf and stem	Antioxidant and LOX-1 inhibitory effects.
Flavonoid and phenolics	Hypericum androsaemum L.	Leaf	Higher radical scavenging and antilipid peroxidation activity
Naringenin; aglycone flavonoid, flavanone	Grapefruits	Fruit	Antilipid peroxidation
(+)-catechin, (-)-epicatechin, and procyanidin B2	Tamarindus indica	Seed	Antioxidant and antilipid peroxidation
Quercetin, kaempferol, myricetin, rutin, naringenin, catechin, fisetin, and gossypetin (flavonoids)	Citrullus colocynthis, Kelussia odoratissima Mozaff., and Carum copticum L.	Leaf and seed	Antilipid peroxidation
Thymol and carvacrol	Thymus vulgaris		Antilipid peroxidation
Ceramicine B (limonoid)	Chisocheton ceramicus	Leaf	Antilipid droplets accumulation activity
(6)-paradol phenolic constituent	Ginger Zingiber officinale	Stem and bud	Antilipid peroxidation
Lycopene	Lycopersicum species	Fruit	Decrease cardiovascular risks
Ginsenoside Rb1 and Rb2	Cudrania cochinchinensis		Antilipid peroxilative effect
Naphthalenic compounds 6-methoxysorigenin and its glycosides, acylates	Rhamnus nakaharai	Leaves	Antilipid peroxidation assay
Madecassoside (terpenoids)	Centella asiatica	Leaves	Antilipid peroxidation
Thiobarbituric acid	Atalantia ceylanica	Leaves	Antilipid peroxidation
Miroestrol (phytoestrogen)	Pueraria mirifica	Tuberous root	Antilipid peroxidation

(Contd...)

Table 1: (Continued)						
Plant natural product	Plant species	Plant part	Biological effects			
Benzenoids, tournefolal, tournefolic acids A and B, and B ethyl ester, and salvianolic acid A, and rosmarinic acid	Tournefortia sarmentosa	Stems	Anti-LDL-peroxidative activity			
quercetin rutinoside (rutin)	Capparis sicula, Capparis orientalis	Aerial parts	Antilipidemic activity			
Methyl ferulate, quercetin 3,4'-dimethyl ether	Terminalia chebula		Antilipid peroxidation activity			
Hederagenin monodesmosides, kalopanaxsaponin A (KPS-A), and sapindoside C	Kalopanax pictus	Stem barks	Antilipid peroxidation effects			
Abietane and seco-abietane diterpenes	Salvia candelabrum	Stem and leaves	Antilipid peroxidation			
Curcurbitacins A, B, C, D, E, I, J, K, and L and Colocynthosides A, and B	Citrullus colocynthis	Fruit and leaves	Anti-inflammatory and antilipidemic			
Phthalides and steroids	Centaurium erythrea L. fam.	Stem and leaves	Antilipid peroxidation			
Total phenolic and flavonoid contents	Aloe barbadensis Miller.	Fleshy leaves	Antilipid peroxidation			

level in rat fed with hypercholesterolemia diet^[103] [Table 1, Figures 3 and 4].

Bel (Aegle Marmelos)

A. marmelos leaf extract shows antihyperlipidemic activity in rats in a dose-dependent manner. It causes a significant reduction in total cholesterol, triglycerides, low-density lipoprotein, very low-density lipoprotein, and significantly increased high-density lipoprotein life. [104] It's unripe fruits also shows antidyslipidemic effect against ISO-induced cardiac stressed rats and restore ISO-induced myocardial infarction [Figures 2 and 3] life. [105] A. marmelos leaf extract pre-treatment increased the activity of Na(+)K(+) ATPase and decreased the activity of Ca(2+) ATPase in the heart and aorta simultaneously [106] [Table 2].

Onion (Allium Cepa)

Onion (*Allium cepa* L.) belongs to family: Liliaceae (lilies). It is an easily digestible aromatic vegetable which is used throughout the world. Onions contain phenolics and flavonoids that have potential anticholesterol and antioxidant properties. Onion contains numerous sulfur compounds including thiosulfinates and thiosulfonates; cepaenes; S-oxides; S, S-dioxides; mono-, di-, and trisulfides; and sulfoxides. Onion contains sulfur compounds, including S-methyl cysteine sulfoxide and allyl propyl disulfide showed hypolipidemic effects life. [107] These compounds also showed anti-atherosclerotic and antithrombotic effects in experimental animal's life. [108] Bioactive compounds found in onion play an important role in the prevention of cardiovascular diseases and cancer [Table 1].

Kareel (C. decidua)

Capparis decidua (CD) is a xerophytic or desert dominating shrub found in desert region of Rajasthan showing strong climatic adaptations. It is a densely branched, thorny plant with smaller scanty and caduceus leaves having pink to red flowers and green berry fruits. Its fruit and shoot extract possesses very strong hypolipidemic efficacy and significantly cut down human plasma triglycerides, total lipids, and phospholipids [Table 1].^[109-111] Plant fruit shows antiatherosclerotic effect^[112] (and its regular use can restore cardiovascular activity).^[113]

Toddalia aculeata

Toddalia aculeata is an important medicinal plant widely used for the treatment of several diseases and disorders. Plant shows antihyperlipidemic activities in high-fat diet fed hyperlipidemic rats.^[114] Plant contains aculeatin that causes lipolysis of 3T3-L1 adipocytes^[115] and assists in restoration of cardiovascular system.^[116] Plant also acts as smooth muscle relaxant^[117] [Table 1].

Ocimum sanctum

Tulsi (*Ocimum sanctum* Linn.) is an aromatic plant belongs to family Lamiaceae. It is traditionally used for the preparation of various Ayurvedic formulations for the treatment of cough, cold, bronchitis, influenza, and asthma. Tulsi leaves show lipid-lowering and antioxidative activities.^[118] It has also been shown to counter metabolic stress does normalization of blood glucose, blood pressure, and lipid levels. It assists in relieving from psychological stress and exerts positive effects

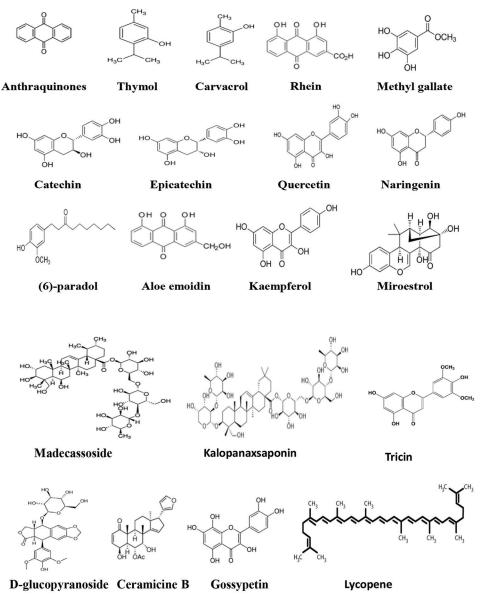


Figure 4: Various antihyperlipidemic bio-organic compounds isolated from various plant species

on memory and cognitive function through its anxiolytic and antidepressant properties.^[119]

OS, commonly known as Holy basil/Tulsi, has been traditionally used to treat cardiovascular diseases (CVD) and manage general cardiac health. OS leaves significantly change the blood lipid profile after a dose 1 g for 4 weeks in albino rabbit. [120] This resulted in significant lowering in serum total cholesterol, triglyceride, phospholipid, and LDL cholesterol levels and significant increase in the HDL cholesterol and total fecal sterol contents. OS contains phenolic compounds and eugenol (EUG) which are traditionally used for treating CVD. [121] Tulsi (OS) polyphenolic extracts were found to have the inherent capacity to inhibit the transcriptional expression of genes, that is, LDLR, LXR alpha, PPARs (alpha, gamma), CD-36, and c-myc which control lipid metabolism, cytokine production, and cellular activity within the arterial wall [Table 1] [Figures 3 and 4].

ANTILIPIDEMIC PLANT NATURAL PRODUCTS

Anthraquinones and Sulfur Compounds

Anthraquinones isolated in ethanolic fractions of total *Radix Rhizoma* Rhei significantly reduce blood levels of total cholesterol, triglycerides, HDL, and LDL.^[122] Gallic acid, a trihydroxybenzoic acid, also demonstrates robust anti-obesity capabilities.^[123] Utilization of niacin and ergosterol may prevent the hypercholesterolemia and incidence of coronary heart diseases [Table 1].^[124] Similar anti-hypercholesterolemic effect is also reported in N-acetylcysteine (NAC) and sesame oil administration in diet-induced hypercholesterolemic mice.^[125] Five major compounds methyl ferulate, methyl p-hydroxycinnamate, p-hydroxybenzaldehyde, tricin, and quercetin 3,4'-dimethyl ether were isolated from EtOAc

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Table 2: Antihyperlipidemic activity in solvent extracts from various plant species				
Scientific name	Plant common name	Plant part extracts/ compound	Biological activity	
Gouania longipetala		Methanol leaf extract	Antilipidemic activities	
Citrullus colocynthis (L.) Schrad.		Leaf extract	Antilipidemic activities	
Symplocos cochinchinensis			Antidiabetic and antilipidemic activities	
Edible oil produced from Punica granatum L.		Seeds		
Momordica charantia		Fruit extract Linn.	Significant lowering of cholesterol and triglyceride levels while elevating HDL cholesterol levels	
Ficus carica		leaves	Antilipidemic and antidiabetic effects of ficusin isolated from	
Capparis orientalis		Stem	These rutin-rich extracts exhibited pronounced dose-dependent enzyme inhibitory activities toward pancreatic lipase.	
Aegle marmelos leaf extract			Shows hyperlipidemic activity in rats dose-dependent reduction of total cholesterol triglycerides, LDL, VLDL, and significantly increased HDL	
Garlic		And cut down lipid contents in experimental animals	Products showed positive effects on lipid metabolism in cholesterol-fed rats	
Bipolaris sp. Bipolaris sp., and Phoma sp. also exhibited significantly		Higher flavonoid and phenolic contents	Exhibited significantly higher radical scavenging and antilipid peroxidation activity than the other isolates	
Wild garlic		Allium ursinum	Wild garlic (<i>Allium ursinum</i>) causes decrease hepatocyte cholesterol synthesis <i>in vitro</i>	
Padina boergesenii		Plant extract	New secondary metabolite, padinolic acid	
Piper trioicum Roxb. and Physalis minima L. extracts			Antilipid peroxidation activity of	
Hypericum androsaemum L.			content of phenolics and flavonoids	
Rhizoma rhei anthraquinones, total Radix et Rhizoma Rhei could		Anthraquinones	Significantly reduce blood levels of total cholesterol, triglycerides, HDL, and LDL	
Phyllanthus emblica leaves		Methanol extract of		
S. cochinchinensis bark		Antilipidemic activities of	Methanolic extract (SCBe) in streptozotocin (STZ)-induced diabetic rats	
Ginger			Lipid peroxidation chemopreventive potential of (6)-paradol, a pungent phenolic constituent of	
<i>Premna integrifolia</i> Linn. Mant. (Verbanacea)		Roots of	Antilipid peroxidation	
		Berberine, suppression of CYP1A expression, and lipid peroxidation by	Beta-naphthoflavone a major isoquinoline alkaloid in herbal plants induced	
Terminalia arjuna (Roxb.) Wight and Arn, and Curcuma longa Linn. extracts			Antilipid-peroxidative effects on serum and platelets <i>in vitro</i>	
Terminalia chebula Retz.		Fruit	Antilipid peroxidative	

(Contd...)

Table 2: (Continued)				
Scientific name	Plant common name	Plant part extracts/ compound	Biological activity	
Cudrania tricuspidata		Root bark flavonoids	Antilipid peroxidation efficacy on human low-density lipoprotein	
Tomato		Tomato paste	Antilipid peroxidative	
Atalantia ceylanica		Plant extract	Antilipid peroxidation	
Chisocheton ceramicus			Ceramicine B, a limonoid with antilipid droplets accumulation activity from	
Apocynum venetum		Leaves extract of	Antioxidant and antilipid peroxidation effects	
Rhamnus alaternus			Antilipid peroxidation	
Premna integrifolia Linn. Mant.		Roots	Antilipid peroxidation	
Agrimonia pilosa			Natural antioxidant sources, prevent oxidation-related diseases	
O. basilicum		Leaves	Antilipid peroxidation effects, potent antioxidant effects	
Sida acuta Burm. f.			Oxygen species/nitrogen species	
Jasminum grandiflorum Linn.			Chemopreventive efficacy and antilipid-peroxidative potential	
Rosa davurica Pall.		Fruit	Prevent the lipid peroxidation	
Pueraria thunbergiana			Antilipid-peroxidative effect	
Kalopanax pictus		MeOH extract of stem barks	Antilipid-peroxidative effect	
Salacia oblonga Wall.			Hypoglycemic and antioxidant activity	
Phyllanthus urinaria may			Antilipid peroxidation	

fraction of *Sorghum bicolor* stem [Table 1]. All five compounds showed antilipid peroxidation activity (IC₅₀ values of 0.5, 0.4, 0.3, and 0.3 microM, respectively). [126] Two compounds kaikasaponin III and tectorigenin isolated from *Pueraria thunbergiana* (Leguminosae) flowers showed potent antilipid-peroxidative effect. [127] Guar gum shows antihypercholesterolemia, antihyperglycemia, and antiobesity activity. [128]

Mozaff. and *Carum copticum* L. contain thymol and carvacrol which show antilipidemic effects^[137] Ceramicine B, a limonoid isolated from *Chisocheton ceramicus*, showed antilipid droplets accumulation activity.^[138] (6)-*paradol* a pungent phenolic constituent of ginger shows antilipid peroxidation and chemopreventive potential.^[139] Similarly, oligostilbenes isolated from the leaf and stem of *Vitis amurensis* potential antioxidant and LOX-1 inhibitory effects [Table 1].^[140]

FLAVONOID AND PHENOLIC COMPOUNDS

Flavonoid and phenolic compounds found in *Hypericum androsaemum* L. (Hypericaceae) and *Bipolaris* sp. show significantlyhigherradicalscavengingand antilipid peroxidation activity. [129,130] Naringenin (NRG), the aglycone flavonoid found in grapefruits, shows antilipid peroxidation activity. [131] Antioxidantand antilipid peroxidation activities are also reported in (+)-catechin, (-)-epicatechin, and procyanidin B2, [132] berberine. [133] D-threo-guaiacylglycerol 8-O-beta-D-(6'-O-galloyl)glucopyranoside, 9-methoxy-D-threo-guaiacylglycerol 8-O-beta-D-(6'-O-galloyl)glucopyranoside, 6''-O-galloyl salidroside, methyl gallate, and quercetin. [134] Plant flavonoids such as quercetin, kaempferol, myricetin, rutin, naringenin, catechin, fisetin, and gossypetin showed antiatherosclerotic effects. [135] *Citrullus colocynthis* [136] and *Kelussia odoratissima*

LYCOPENE

Lycopene is known to decrease cardiovascular risks. Madecassoside also shows protective effect against myocardial ischemia reperfusion injury and effects of antilipid peroxidation.[141] GSTC3 decreased serum total cholesterol, low-density lipoprotein (LDL), very low-density lipoprotein (VLDL), triglycerides, and phospholipids at a dose of 100 mg/kg body weight.^[29] It causes hypolipidemic effect due to partial inhibition of HMG-CoA reductase activity.[29] Ginsenoside Rb1 and Rg1 isolated from Cudrania cochinchinensis var. gerontogea, [142] naphthalenic compounds 6-methoxysorigenin, and its glycosides [i.e., 6-methoxysorigenin-8-O-glucoside, alpha-sorinin, 6-methoxysorigenin-8-rutinoside], two acylates (peracetate and perpropionate) from Rhamnus nakaharai showed antilipid peroxidation activity.[143] Madecassoside (MA) one of the principle terpenoids in Centella asiatica shows protective effects against myocardial ischemia-reperfusion injury antilipid peroxidation.[144] Atalantia cevlanica leaves contain thiobarbituric acid showed antilipid peroxidation.^[145] Miroestrol (MR) is a potent phytoestrogen from the tuberous root of Pueraria mirifica, antilipid peroxidation; and normalizes the superoxide dismutase (SOD), catalase (CAT), and glutathione peroxidase activity on glutathione content in the livers and uteri of β-naphthoflavone (BNF)-treated mice.[145] Anti-LDL-peroxidative activity was detected in benzenoids, tournefolal, tournefolic acids A and B, and B ethyl ester, together with salvianolic acid A, isosalvianolic acid C, lithospermic acid, salvianolic acid F, and rosmarinic acid isolated from the stems of Tournefortia sarmentosa. [146] Capparis sicula subsp. sicula and Capparis orientalis aerial parts showed antilipidemic activity due to the presence of quercetin rutinoside (rutin).[147] Terminalia chebula RETZ contains methyl ferulate^[148] and Kalopanax pictus stem barks contain two hederagenin monodesmosides, kalopanaxsaponin A (KPS-A) and sapindoside C, which exhibit significant antilipid peroxidation effects.[149] Salvia candelabrum shows antilipid peroxidation activity due to the presence of abietane and seco-abietane diterpenes [Table 1].[150]

CARDIOPROTECTIVE EFFECTS

Bel is used for the preparation of various Ayurvedic traditional medicines. It has wide ethnomedicinal uses because of its strong cardioprotective activity.^[151] The leaf extract of A. marmelos shows preventive effect in isoprenaline isoproterenol (ISO)-induced myocardial infarction in rats.[106] It restores creatine kinase and lactate dehydrogenase activity in the heart of isoprenaline-treated rats. Bel is also used as a cardiac depressant[152-154] and found biologically active against several major diseases including cancer, diabetes, and cardiovascular diseases.[155] Its leaf extract shows antihyperglycemic and antihyperlipidemic activities and minimizes the cardiovascular risk factors associated with diabetes.^[156] Lanosteryl triterpenes isolated from Protorhus longifolia act as a cardioprotective agent in diabetic cardiomyopathy and ischemic heart diseases.[157] Periplogenin-3-O–Dglucopyranosyl-(1→6)-Dglucopyaranosyl-(1→4)-Dcymaropyranoside, isolated from A. marmelos, shows protective effects against DOX-induced cardiovascular morbidities and hepatotoxicity in rats.[158] Plant polyphenols are also used for the prevention of heart diseases.[159] Garlic-derived allicin shows cardiovascular benefits and antioxidant properties.[160] It also shows significant anti-atherosclerotic potential and unique vascular protective properties.[161] Both processing and cooking conditions of Allium sp. induced antiplatelet activity and thiosulfinate content.^[162]

Raw garlic and its essential oil show serum fibrinolytic activity and anti-clotting effects that could become a solution of cardiovascular diseases^[163] for patients suffering from coronary artery disease. [46] The long-term use of garlic causes significant improvement in ischemic heart diseases. Both raw

garlic and fried garlic improve microhemovascular system^[164] and increase fibrinolytic activity in patients if continued therapy is provided [Table 1, Figures 3 and 4]. [165] Garlic shows endothelium mediated vasorelaxant response in isolated rat aorta which tone up heart physiology. [166] Garlic may act on the nitric oxide system and exert effects on the elastic properties of vasculature. [166] It restores systemic blood pressure [167a,b] and shows protective effect in aorta of the elderly patients. [168] It may be endothelium dependent and independent. Garlic powder put positive effect on cutaneous microcirculation of diseased [167a] and healthy persons. [167b] Similarly, dried ethanol-water extract of garlic shows acute effect on the microhemovascular system of the skin [167a,b] and restores function of conjunctival vessels [168] [Table 1, Figures 3 and 4].

However, potent activation of nitric oxide synthase by garlic may be highly useful in multiple therapeutic applications.[169] Both allicin and ajoene, compounds derived from garlic, induce nitric oxide synthase system^[170] that prevents hypertension.[171] Garlic does pulmonary vasorelaxation due to the presence of allicin.[172] Similarly, aqueous garlic extract shows beneficial effect on the blood vascular system of streptozotocin diabetic rats.[173,174] Onions bulbs contain flavonoids which provide relief in heartburn^[175] cardiovascular problems.[176] Onion (Allium cepa) leaves also showed cardioprotective and antioxidant activity in doxorubicin (DOX)-induced cardiotoxicity in rats.[176,177] Its use can restore and control ambulatory blood pressure and endothelial function in overweight-to-obese patients affected with sever hypertension.[178] Onion extract and onion soup showed inhibitory activity toward platelet aggregation. [179-182] This activity is due presence of quercetin, in green onion prohibit bulbs.[183,184] However, the use of green onion in salads successfully decreases blood pressure in hypertensive subjects.[178] It extends immense cardiovascular benefits in human subjects. Thiosulfinates dimethyl- and diphenylthiosulfinate slow down thrombocyte biosynthesis. [181,182] Cynodon dactylon significantly decreased the total cholesterol, low-density lipoprotein, and triglyceride levels in blood serum of alloxan-induced diabetic rats^[185] [Figures 3 and 4].

Gingko biloba extract (EGb) shows cardioprotective protective effect against myocardial impairment in diabetic rats.[186] It assists in repairing (EGb) of damages induced by lysophosphatidylcholine in vascular endothelial cells in vitro.[187] Similar cardioprotective potential is also found in few other plant species, that is, Rauvolfia serpentina, Terminalia arjuna, Coriandrum sativum, Elettaria cardamomum, Piper nigrum, Allium sativum, Crataegus oxyacantha, [188] and Sida rhombifolia (L.).[189] In addition, cardioprotective effects are also reported in puerarin isolated from Pueraria lobata Ohwi., and Danshensu isolated from the Chinese herb, Salvia miltiorrhiza. These bio-organics are used for the treatment of acute ischemic myocardial injury ischemic heart disease. [190] Addition of dietary fibers decreases serum cholesterol, triglycerides, increasing the high-density lipoprotein cholesterol level, and the management of glycemic indices and obesity.^[128] Gum guggul is used as a therapeutic agent in the treatment of hyperlipidemia and associated cardiac disorders such as hypertension and ischemia.^[191]

At global level, there is a rapid increase which has been observed in coronary heart disease (CHD) and cardiovascular diseases. This global increase in cases of hyperlipidemia is due to unhealthy eating habits, obesity, and physical inactivity.[192] In India, 17% of total deaths occurred were due to hyperlipidemia. Few lipid-soluble drugs inhibit the enzymes responsible for the synthesis of cholesterol and cut down its level significantly.[193] It could be contributed by antihyperlipidemic natural plant products, that is, acetone extract of the fruits of Capsicum frutescens. [194] Ethanolic extract of C. tamala leaves is showed hypoglycemic and antihyperglycemic activities.[195] Administration of C. tamala leaves extract in diabetic rats restored the level of blood glucose to near normal levels. In addition, total cholesterol, TG, LDL, and VLDL cholesterol levels in diabetic rats were a significantly increased.^[196] Regular dietary use of C. tamala leaves can cure diabetes associated with hyperlipidemia. No doubt leaf extract promotes the insulin release from the undestroyed β-cells.^[197] Cinnamomum verum shows positive cardioprotective effects and reduces blood pressure, plasma glucose, obesity, and ameliorating dvslipidemia. Cinnamomum cassia extracts are also used to cure gastritis, diabetes, blood circulation disturbance, and inflammatory diseases [Figures 3 and 4].[198] C. cassia contains coumarin, trans-cinnamic acid, cinnamaldehyde, 2-hydroxycinnamaldehyde, 2-methoxycinnamaldehyde (6), 2-hydroxy-cinnamylalcohol, benzoic acid, (+)-syringaresinol, and phenethyl (E)-3-[4-methoxyphenyl]-2-propenoate bioorganic compounds with antihyperlipidemic activity. [199]

CONCLUSION

There are thousands of plant species which possess antioxidants that could be used for antihyperlipidemic drug formulations. Before use herbal preparations, their safety, potency, toxicity, life-threatening adverse effects, and possible herb-drug interactions must be evaluated, analyzed, and critically discussed. In particular, before use of any herbal drug formulation, there is immense need to determine clinical and therapeutic dose level. Its efficacy and side effects by testing them in 2-fold blind placebo experiments in animal models. For making public health index, conventional risk prediction algorithms must prepare to identify intensive risk factors, microlevel changes in various biomolecules, function of enzymes, and their metabolic pathways. For more appropriate judgment of CVDs, there is a need to check lipid profile, triglyceride to HDLp (high-density lipoproteins) ratio, lipoprotein cholesterol ratio, lipid-lipoprotein ratio, LDL cholesterol level, HDLp and apolipoprotein levels, lipoprotein and LTPs ratio, sphingolipids, omega-3 index, and ST2 level. However, major cardiovascular risks can be identified by measurement of non-HDL cholesterol, total HDL cholesterol and triglyceride, and HDL cholesterol ratios. There is also an urgent need of new more authentic, appropriate, and reliable diagnostic and therapeutic markers to confirm CVDs, well in time to start the clinical aid to the patients. Well-designed clinical trials are required to test action of herbal drugs on tissues, cells, and organ system of patient to lower down usage specific adverse effects.

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REFERENCES

- Shi F, Reuss L, Wang Y. Potential of natural products in the inhibition of adipogenesis through regulation of PPARγ expression and/or its transcriptional activity. Molecules 2016;21:1278.
- Abdul HP, Khan JA, Ullah I, Ullah S. Synergistic combinatorial antihyperlipidemic study of selected natural antioxidants; modulatory effects on lipid profile and endogenous antioxidants. Lipids Health Dis 2016;15:151.
- 3. Yan B, Ren J, Zhang Q, Gao R, Zhao F, Wu J, *et al.* Antioxidative effects of natural products on diabetic cardiomyopathy. J Diabetes Res 2017;2017:2070178.
- 4. Lloyd-Jones D, Adams RJ, Brown TM, Carnethon M, Dai S, De Simone G, *et al.* Heart disease and stroke statistics-2010 update: A report from the American heart association. Circulation 2010;7:948-54.
- 5. Lusis AJ. Atherosclerosis. Nature 2000;407:233-41.
- Joshi SC. Plant and plant products used as hypolipidaemic/ antiatherosclerotic agents: An overview. Proc Zool Soc India 2005;4:27-32.
- 7. Kirichenko TV, Sukhorukov VN, Markin AM, Nikiforov NG, Liu PY, Sobenin IA, *et al.* Medicinal plants as a potential and successful treatment option in the context of atherosclerosis. Front Pharmacol 2020;11:403.
- 8. Souza TD, Mengi S, Hassarajani AS, Chattopadhayay S. Efficacy study of the bioactive fraction(F-3) of *Acorus calamus* in hyperlipidemia. Indian J Pharmacol 2007;39:196-200.
- Alexander NO, Sobenin IA, Revin VV, Bobryshev YV. Development of antiatherosclerotic drugs on the basis of natural products using cell model approach. Oxid Med Cell Longev 2015;2015:463797.
- Ghosal S, Bhattacharya A, Bhattacharya SK. Antioxidant effects of *Emblica officinalis* and their biological consequences. In: Majumdar DK, Govil JN, Singh VK, editors. Recent Progress in Medicinal Plants. Vol. 8. New Delhi: Studium Press; 2003.
- 11. Kris-Etherton PM, Hecker KD, Bonanome A, Coval SM, Binkoski AE, Hilpert KF, *et al.* Bioactive compounds in foods: Their role in the prevention of cardiovascular

- disease and cancer. Am J Med 2002:113:71S-88S.
- 12. Serafini M, Bellocco R, Wolk A, Ekstrom AM. Total antioxidant potential of fruit and vegetables and risk of gastric cancer. Gastroenterology 2002;123:985-91.
- Blazovics A, Lugasi A, Hagymasi K, Szentmihalyi K, Kery A. Natural antioxidants and tissue regenerations: Curative effect and reaction mechanism. In: Majumdar DK, Govil JN, Singh VK, editors. Recent Progress in Medicinal Plants. New Delhi: Studium Press; 2003.
- 14. Mahmood ZA, Sualeh M, Mahmood SBZ, Karim MA. Herbal treatment for cardiovascular disease the evidence based therapy. Pak J Pharm Sci 2010;23:119-24.
- 15. Shaito A, Thuan DT, Phu HT, Nguyen TH, Hasan H, Halabi S, *et al.* Gianfranco pintus herbal medicine for cardiovascular diseases: Efficacy, mechanisms, and safety. Front Pharmacol 2020;11:422.
- 16. Robin D. The International Cocoa Trade. Sawston: Woodhead Publishing; 1999. p. 169.
- 17. Thomas A. Fats and Fatty Oils. Ullmann's Encyclopedia of Industrial Chemistry. Weinheim: Wiley-VCH; 2002.
- Yara-Varón E, Li Y, Balcells M, Canela-Garayoa R, Fabiano-Tixier AS, Chemat F. Vegetable oils as alternative solvents for green oleo-extraction. Purif Form Food Nat Prod Mol 2017;22:1474.
- 19. Lautié E, Russo O, Ducrot P, Boutin JA. Unraveling plant natural chemical diversity for drug discovery purposes. Front Pharmacol 2020;11:397.
- Ramachandra PH. Lipid peroxidation in culinary oils subjected to thermal stress. Indian J Clin Biochem 2000;15:1-5.
- Tańska M, Roszkowska B, Skrajda M, Dąbrowski G. Commercial cold pressed flaxseed oils quality and oxidative stability at the beginning and the end of their shelf life. J Oleo Sci 2016;65:111-21.
- 22. Fursule RA, Patil SD. Hepatoprotective and antioxidant activity of *Phaseolus trilobus*, Ait on bile duct ligation induced liver fibrosis in rats J Ethnopharmacol 2010;129:416-9.
- 23. Zheng J, Zhou Y, Li S, Zhang P, Zhou T, Xu DP, Hua-Bin L. Effects and mechanisms of fruit and vegetable juices on cardiovascular diseases. Int J Mol Sci 2017;18: 555.
- 24. Tariq S, Imran M, Mushtaq Z, Asghar N. Phytopreventive antihypercholesterolmic and antilipidemic perspectives of zedoary (*Curcuma zedoaria* Roscoe.) herbal tea Lipids Health Dis 2016;15:39.
- 25. Hosseini S, Nili-Ahmadabadi A, Nachvak SM, Dastan D, Moradi S, Abdollahzad H, et al. Antihyperlipidemic and antioxidative properties of Pistacia atlantica subsp. kurdica in streptozotocin-induced diabetic mice. Diabetes Metab Syndr Obes 2020;13:1231-6.
- 26. Ku CS, Kim B, Pham TX, Yang Y, Weller CL, Carr TP, et al. Hypolipidemic effect of a blue-green alga (nostoc commune) is attributed to its nonlipid fraction by decreasing intestinal cholesterol absorption in C57BL/6J mice. J Med Food 2015;18:1214-22.
- 27. Aloulou A, Hamden K, Elloumi D, Ali MB, Hargafi K, Jaouadi B, *et al.* Hypoglycemic and antilipidemic properties of kombucha tea in alloxan-induced diabetic

- rats. BMC Complement Altern Med 2012;12:63.
- 28. Fan W, Fan L, Peng C, Zhang Q, Wang L, Li L, *et al.* Traditional uses, botany, phytochemistry, pharmacology, pharmacokinetics and toxicology of *Xanthium strumarium* L.: A review. Molecules 2019;24:359.
- Manalil JJ, Baby M, Ramavarma SK, Suseela IM, Padikkala J, Raghavamenon AJ. Development of an anti-atherosclerotic polyherbal formulation: GSTC3. Environ Pathol Toxicol Oncol 2015;34:237-48.
- 30. Barter PJ, Rye KA. New era of lipid-lowering drugs. Pharmacol Rev 2016;68:458-75.
- 31. Rouhi-Boroujeni H, Rouhi-Boroujeni H, Gharipour M, Mohammadizadeh F, Ahmadi S, Rafieian-Kopaei M. Systematic review on safety and drug interaction of herbal therapy in hyperlipidemia: A guide for internist. Acta Biomed 2015;86:130-6.
- 32. Asghar N, Mushtaq Z, Arshad MU, Imran M, Ahmad RS, Hussain SM. Phytochemical composition, antilipidemic and antihypercholestrolemic perspectives of Bael leaf extracts. Lipids Health Dis 2018;17:68.
- 33. Olorunnisola OS, Adetutu A, Afolayan AJ, Owoade AO. Effect of methanolic leaf extract of *Talinum triangulare* (Jacq). Willd. on biochemical parameters in diet induced dyslipidemia wistar rats. Pharmacogn Mag 2016;12:333-9.
- 34. Fernandes NP, Lagishetty CV, Panda VS, Naik SR. An experimental evaluation of the antidiabetic and antilipidemic properties of a standardized *Momordica charantia* fruit extract. BMC Complement Altern Med 2007;7:29.
- 35. Moniruzzaman M, Rokeya B, Ahmed S, Bhowmik A, Khalil I, Gan SH. *In vitro* antioxidant effects of *Aloe barbadensis* miller extracts and the potential role of these extracts as antidiabetic and antilipidemic agents on streptozotocin-induced Type 2 diabetic model rats. Molecules 2012;17:12851-67.
- 36. Marrelli M, Morrone F, Argentieri MP, Gambacorta L, Conforti F, Avato P. Phytochemical and biological profile of *Moricandia arvensis* (L.) DC. An inhibitor of pancreatic lipase. Molecules 2018;23:2829.
- 37. Ali L, Khan AL, Al-Broumi M, Al-Harrasi R, Al-Kharusi L, Hussain J, *et al*. New enzyme-inhibitory triterpenoid from marine macro brown alga *Padina boergesenii* Allender and Kraft. Mar Drugs 2017;15:19.
- 38. Shabbir M, Khan MR, Saeed N. Assessment of phytochemicals, antioxidant, anti-lipid peroxidation and anti-hemolytic activity of extract and various fractions of *Maytenus royleanus* leaves. BMC Complement Altern Med 2013;13:143.
- 39. Tahir I, Khan MR, Shah MA, Aftab M. Evaluation of phytochemicals, antioxidant activity and amelioration of pulmonary fibrosis with *Phyllanthus emblica* leaves. BMC Complement Altern Med 2016;16:406.
- 40. Mkadmini KH, Hammami M, Rihouey C, Le Cerf D, Ksouri R, Majdoub H. Optimization extraction of polysaccharide from Tunisian *Zizyphus lotus* fruit by response surface methodology: Composition and antioxidant activity. Food Chem 2016;212:476-84.

- 41. Bag A, Bhattacharyya SK, Chattopadhyay RR. The development of *Terminalia chebula* Retz. (*Combretaceae*) in clinical research. Asian Pac J Trop Biomed 2013;3:244-52.
- 42. Ghulam A, Shahzad M, Saddiqe Z, Hassan MJ, Saba S, Rafique J, *et al.* Various fractions of *Hypericum x moserianum* and *Hypericum ericoides* possess antiglycation, anti-lipid peroxidation, antioxidative activities and non-toxic effects *in vitro*. Pak J Pharm Sci 2015;28:933-8.
- 43. Arunachalam K, Parimelazhagan T. Antidiabetic activity of *Ficus amplissima* Smith, Bark extract in streptozotocin induced diabetic rats. J Ethnopharmacol 2013;147:302-10.
- 44. Choi ES, Kim JS, Kwon KH, Kim HS, Cho NP, Cho SD. Methanol extract of *Sanguisorba officinalis* L. with cytotoxic activity against PC3 human prostate cancer cells. Mol Med Rep 2012;6:670-4.
- 45. Yu T, Lee YJ, Yang HM, Han S, Kim JH, Lee Y, *et al.* Inhibitory effect of *Sanguisorba officinalis* ethanol extracts on NO and PGE₂ production is mediated by suppression of NF-κB and AP-1 activation signaling cascade. J Ethnopharmacol 2011;134:11-7.
- 46. Lau YS, Kwan CY, Ku TC, Hsieh WT, Wang HD, Nishibe S, *et al. Apocynum venetum* leaf extract, an antihypertensive herb, inhibits rat aortic contraction induced by angiotensin II: A nitric oxide and superoxide connection. J Ethnopharmacol 2012;143:565-71.
- 47. Dinakaran SK, Saraswathi NR, Nalini VR, Srisudharson, Bodanapu VR, Avasarala H, *et al*. Anti lipid peroxidation activity of *Piper trioicum* Roxb. and *Physalis minima* L. extracts. Pak J Pharm Sci 2011;24:411-3.
- 48. Ammar RB, Neffati A, Skandrani I, Sghaier MB, Bhouri W, Ghedira K, *et al.* Anti-lipid peroxidation and induction of apoptosis in the erythroleukaemic cell line K562 by extracts from (Tunisian) *Rhamnus alaternus* L. (*Rhamnaceae*). Nat Prod Res 2011;25:1047-58.
- 49. Meera R, Devi P, Kameswari B, Madhumitha B, Merlin NJ Antioxidant and hepatoprotective activities of *Ocimum basilicum* Linn. and *Trigonella foenum-graecum* Linn. against H₂O₂ and CCL₄ induced hepatotoxicity in goat liver. Indian J Exp Biol 2009;47:584-90.
- 50. Visavadiya NP, Soni B, Soni B, Madamwar D. Suppression of reactive oxygen species and nitric oxide by *Asparagus racemosus* root extract using *in vitro* studies. Cell Mol Biol (Noisy-le-Grand) 2009;55:OL1083-95.
- 51. Gokani RH, Lahiri SK, Santani DD, Shah MB. Evaluation of anti-inflammatory and antioxidant activity of *Premna integrifolia* root. J Complement Integr Med 2011;8:3840.
- 52. Sathaye S, Bagul Y, Gupta S, Kaur H, Redkar R. Hepatoprotective effects of aqueous leaf extract and crude isolates of *Murraya koenigii* against *in vitro* ethanol-induced hepatotoxicity model. Exp Toxicol Pathol 2011;63:587-91.
- 53. Nwanjo HU, Okafor MC, Oze GO. Anti-lipid peroxidative activity of *Gongronema latifolium* in

- streptozotocin-induced diabetic rats. Niger J Physiol Sci 2006;21:61-5.
- 54. Kolanjiappan K, Manoharan S. Chemopreventive efficacy and anti-lipid peroxidative potential of *Jasminum grandiflorum* Linn. on 7,12-dimethylbenz(a) anthracene-induced rat mammary carcinogenesis. Fundam Clin Pharmacol 2005;19:687-93.
- 55. Bhoopat L, Srichairatanakool S, Kanjanapothi D, Taesotikul T, Thananchai H, Bhoopat T. Hepatoprotective effects of lychee (*Litchi chinensis* Sonn.): A combination of antioxidant and anti-apoptotic activities. J Ethnopharmacol 2011;136:55-66.
- 56. Jiao SP, Chen B, Du PG. Anti-lipid peroxidation effect of *Rosa davurica* Pall. fruit. Zhong Xi Yi Jie He Xue Bao 2004:2:364-5.
- 57. Kamalakkannan N, Prince PS. Effect of *Aegle marmelos* Correa. (Bael) fruit extract on tissue antioxidants in streptozotocin diabetic rats. Indian J Exp Biol 2003;41:1285-8.
- 58. Hsu YM, Lai CH, Chang CY, Fan CT, Chen CT, Wu CH. Characterizing the lipid-lowering effects and antioxidant mechanisms of tomato paste. Biosci Biotechnol Biochem 2008;72:677-85.
- 59. Hsu TY, Sheu SC, Liaw ET, Wang TC, Lin CC. Antioxidant activity and effect of *Pinus morrisonicola* Hay on the survival of leukemia cell line U937. Phytomedicine 2005;12:663-9.
- 60. Gayathri V, Asha VV, John JA, Subramoniam A. Protection of immunocompromised mice from fungal infection with a thymus growth-stimulatory component from *Selaginella involvens*, a fern. Immunopharmacol Immunotoxicol 2011;33:351-9.
- 61. Moniruzzaman M, Rokeya B, Ahmed S, Bhowmik A, Khalil MI, Gan SH. *In vitro* antioxidant effects of *Aloe barbadensis* Miller extracts and the potential role of these extracts as antidiabetic and antilipidemic agents on streptozotocin-induced Type 2 diabetic model rats. Molecules 2012;17:12851-67.
- 62. Sunil C, Agastian P, Kumarappan C, Ignacimuthu S. *In vitro* antioxidant, antidiabetic and antilipidemic activities of *Symplocos cochinchinensis* (Lour.) S. Moore bark. Food Chem Toxicol 2012;50:1547-53.
- 63. Pushparaj P, Tan CH, Tan BK. Effects of *Averrhoa bilimbi* leaf extract on blood glucose and lipids in streptozotocindiabetic rats. J Ethnopharmacol 2000;72:69-76.
- 64. Zhou S, Xu C, Zhou N, Huang Y, Huang L, Chen X, et al. Mechanism of protective action of *Phyllanthus urinaria* L. against injuries of liver cells. Zhongguo Zhong Yao Za Zhi 1997;22:109-11.
- 65. Wu H, Kong L, Wu M, Xi P. Effects of different processed products of radix *Angelica sinensis* on clearing out oxygen free radicals and anti-lipid peroxidation. Zhongguo Zhong Yao Za Zhi 1996;21:599-601.
- 66. Hou WC, Hsu FL, Lee MH. Yam. *Dioscorea batatas* tuber mucilage exhibited antioxidant activities *in vitro*. Planta Med 2002;68:1072-6.
- 67. Subashini R, Ragavendran B, Gnanapragasam A, Yogeeta SK, Devaki T. Biochemical study on the

- protective potential of *Nardostachys jatamansi* extract on lipid profile and lipid metabolizing enzymes in doxorubicin intoxicated rats. Pharmazie 2007;62:382-7.
- 68. Li JM, Kong LD. Advances in the study on depressive and anxiety disorders treated with traditional Chinese medicine and herbal drugs. Zhongguo Zhong Yao Za Zhi 2001;26:805-7.
- 69. Wu W, Luo H, Hou C. *In vitro* experiment on scavenging oxy-radical and suppressing myocardial lipid peroxidation with sini decoction Zhongguo Zhong Yao Za Zhi 1995;20:690-1.
- Irudayaraj SS, Stalin A, Sunil C, Duraipandiyan V, Al-Dhabi NA, Ignacimuthu S. Antioxidant, antilipidemic and antidiabetic effects of ficusin with their effects on GLUT4 translocation and PPARγ expression in Type 2 diabetic rats. Chem Biol Interact 2016;256:85-93.
- 71. Aviello G, Abenavoli L, Borrelli F, Capasso R, Izzo AA, Lembo F, *et al.* Garlic: Empiricism or science? Nat Prod Commun 2009;4:1785-96.
- 72. Berginc K, Kristl A. The mechanisms responsible for garlic drug interactions and their *in vivo* relevance. Curr Drug Metab 2013;14:90-101.
- 73. Sabater-Molina M, Larqué E, Torrella F, Zamora S. Dietary fructooligosaccharides and potential benefits on health. J Physiol Biochem 2009;65:315-28.
- 74. Khatua TN, Adela R, Banerjee SK. Garlic and cardioprotection: Insights into the molecular mechanisms. Can J Physiol Pharmacol 2013;91:448-58.
- 75. Keys A. Wine, garlic, and CHD in seven countries. Lancet 1980;1:145-6.
- 76. Campbell JH, Efendy JL, Smith NJ, Campbell GR. Molecular basis by which garlic suppresses atherosclerosis. J Nutr 2001;131:1006S-9.
- 77. Koscielny J, Klüssendorf D, Latza R, Schmitt R, Radtke H, Siegel G, *et al.* The antiatherosclerotic effect of *Allium sativum*. Atherosclerosis 1999;144:237-49.
- 78. Sendl A, Elbl G, Steinke B, Redl K, Breu W, Wagner H. Comparative pharmacological investigations of *Allium ursinum* and *Allium sativum*. Planta Med 1992;58:1-7.
- 79. Ali M. Mechanism by which garlic (*Allium sativum*) inhibits cyclooxygenase activity. Effect of raw versus boiled garlic extract on the synthesis of prostanoids. Prostaglandins Leukot Essent Fatty Acids 1995;53:397-400.
- 80. Arunkumar R, Sharmila G, Elumalai P, Senthilkumar K, Banudevi S, Gunadharini DN, *et al.* Effect of diallyl disulfide on insulin-like growth factor signaling molecules involved in cell survival and proliferation of human prostate cancer cells *in vitro* and *in silico* approach through docking analysis. Phytomedicine 2012;19:912-23.
- 81. Chi MS. Effects of garlic products on lipid metabolism in cholesterol-fed rats (41494). Proc Soc Exp Biol Med 1982;171:174-8.
- 82. Chi MS, Koh ET, Stewart TJ. Effects of garlic on lipid metabolism in rats fed cholesterol or lard. J Nutr 1982;112:241-8.
- 83. Qureshi AA, Abuirmeileh N, Din ZZ, Elson CE, Burger WC. Inhibition of cholesterol and fatty acid

- biosynthesis in liver enzymes and chicken hepatocytes by polar fractions of garlic. Lipids 1983;18:343-8.
- 84. Qureshi AA, Crenshaw TD, Abuirmeileh N, Peterson DM, Elson CE. Influence of minor plant constituents on porcine hepatic lipid metabolism. Impact on serum lipids. Atherosclerosis 1987;64:109-15.
- 85. Gebhardt R, Beck H, Wagner KG. Inhibition of cholesterol biosynthesis by allicin and ajoene in rat hepatocytes and HepG2 cells. Biochim Biophys Acta 1994;1213:57-62.
- 86. Yeh YY, Yeh SM. Garlic reduces plasma lipids by inhibiting hepatic cholesterol and triacylglycerol synthesis. Lipids 1994;29:189-93.
- 87. Gupta N, Porter TD. Garlic and garlic-derived compounds inhibit human squalene monooxygenase. J Nutr 2001;131:1662-7.
- 88. Gebhardt R, Beck H. Differential inhibitory effects of garlic-derived organosulfur compounds on cholesterol biosynthesis in primary rat hepatocyte cultures. Lipids 1996;31:1269-76.
- 89. Liu L, Yeh YY. Inhibition of cholesterol biosynthesis by organosulfur compounds derived from garlic. Lipids 2000;35:197-203.
- 90. Bordia A, Bansal HC. Letter: Essential oil of garlic in prevention of atherosclerosis. Lancet 1973;2:1491-2.
- 91. Sendl A, Schliack M, Löser R, Stanislaus F, Wagner H. Inhibition of cholesterol synthesis *in vitro* by extracts and isolated compounds prepared from garlic and wild garlic. Atherosclerosis 1992;94:79-85.
- 92. Ide N, Lau BH. Garlic compounds protect vascular endothelial cells from oxidized low density lipoprotein-induced injury. J Pharm Pharmacol 1997;49:908-11.
- 93. Efendy JL, Simmons DL, Campbell GR, Campbell JH. The effect of the aged garlic extract, 'Kyolic', on the development of experimental atherosclerosis. Atherosclerosis 1997;132:37-42.
- 94. Jain RC, Konar DB. Effect of garlic oil in experimental cholesterol atherosclerosis. Atherosclerosis 1978;29:125-9.
- 95. Orekhov AN, Tertov VV. *In vitro* effect of garlic powder extract on lipid content in normal and atherosclerotic human aortic cells. Lipids 1997;32:1055-60.
- 96. Kiesewetter H. Long-term effect of garlic powder tablets on the development of plaque formation in the carotid branches of both femoral arteries a preliminary report. Eur J Clin Res 1996;8:34-5.
- 97. Durak Ì, Öztürk HS, Olcay E, Güven C. Effects of garlic extract supplementation on blood lipid and antioxidant parameters and atherosclerotic plaque formation process in cholesterol-fed rabbits. J Herb Pharmacother 2002;2:19-32.
- 98. Phelps S, Harris WS. Garlic supplementation and lipoprotein oxidation susceptibility. Lipids 1993;28:475-7.
- 99. Munday JS, James KA, Fray LM, Kirkwood SW, Thompson KG. Daily supplementation with aged garlic extract, but not raw garlic, protects low density lipoprotein against *in vitro* oxidation. Atherosclerosis 1999;143:399-404.

- 100. Byrne DJ, Neil HA, Vallance DT, Winder AF. A pilot study of garlic consumption shows no significant effect on markers of oxidation or sub-fraction composition of low-density lipoprotein including lipoprotein(a) after allowance for non-compliance and the placebo effect. Clin Chim Acta 1999;285:21-33.
- 101. Dwivedi C, John LM, Schmidt DS, Engineer FN. Effects of oil-soluble organosulfur compounds from garlic on doxorubicin-induced lipid peroxidation. Anticancer Drugs 1998;9:291-4.
- 102. Ide N, Nelson AB, Lau BH. Aged garlic extract and its constituents inhibit Cu(2)-induced oxidative modification of low density lipoprotein. Planta Med 1997;63:263-4.
- 103. Ebrahimi T, Behdad B, Abbasi MA, Rabati RG, Fayyaz AF, Behnod V, *et al.* High doses of garlic extract significantly attenuated the ratio of serum LDL to HDL level in rat-fed with hypercholesterolemia diet. Diagn Pathol 2015;10:74.
- 104. Bhuvaneswari R, Sasikumar K. Antihyperlipidemic activities of *Aegle marmelos* (L) Corr., Leaf extract in Triton WR-1339 induced hyperlipidemic rats. Pharm Glob 2013;3:1-3.
- 105. Krushna G, Kareem MA, Devi KL. Antidyslipidaemic effect of *Aegle marmelos* Linn. Fruit on Isoproterenol induced myocardial injury in rats. Inter J Pharmacol. 2009;6:1-5.
- 106. Prince PS, Rajadurai M. Preventive effect of *Aegle marmelos* leaf extract on isoprenaline-induced myocardial infarction in rats: Biochemical evidence. J Pharm Pharmacol 2005;57:1353-7.
- 107. Kumari K, Augusti KT. Lipid lowering effect of S-methyl cysteine sulfoxide from *Allium cepa* Linn in high cholesterol diet fed rats. J Ethnopharmacol 2007;109:367-71.
- 108. Augusti KT, Arathy SL, Asha R, Ramakrishanan J, Zaira J, Lekha V, et al. A comparative study on the beneficial effects of garlic (Allium sativum Linn), amla (Emblica officinalis Gaertn) and onion (Allium cepa Linn) on the hyperlipidemia induced by butter fat and beef fat in rats. Indian J Exp Biol 2001;39:760-6.
- 109. Sharma I, Gusain D, Sharma A, Dixit VP. Hypolipidaemic effects of *Capparis decidua* fruit extract (50% EtOH) in cholesterol-fed rabbits. Indian Drugs 1991;28:412-6.
- 110. Chahlia N. Effect of *Capparis decidua* on hypolipidemic activity in rats. J Med Plant Res 2009;3:481-4.
- 111. Purohit A, Vyas KB. Hypolipidaemic efficacy of *Capparis deciduas* fruit and shoot extracts in cholesterol fed rabbits. Indian J Exp Biol 2005;43:863-6.
- 112. Goyal R, Grewal RB. The influence of teent (*Capparis decidua*) on human plasma triglycerides, total lipids and phospholipids. Nutr Health 2003;17:71-6.
- 113. Rashid S, Lodhi F, Ahmad M, Usmanghani K. Preliminary cardiovascular activity evaluation of capparidisine, a spermidine alcohol from *Capparis decidua*. Pak J Pharmacol 1989;6;6-16.

- 114. Irudayaraj SS, Sunil C, Duraipandiyan V, Ignacimuthu S. *In vitro* antioxidant and antihyperlipidemic activities of *Toddalia asiatica* (L) Lam. leaves in Triton WR-1339 and high fat diet induced hyperlipidemic rats. Food Chem Toxicol 2013;60:135-40.
- 115. Watanabe A, Kato T, Ito Y, Yoshida I, Harada T, Mishima T, et al. Aculeatin, a coumarin derived from *Toddalia asiatica* (L.) Lam., enhances differentiation and lipolysis of 3T3-L1 adipocytes. Biochem Biophys Res Commun 2014;453:787-92.
- 116. Guo S, Li S, Peng Z, Ren X. Isolation and identification of active constituent of *Toddalia asiatica* in cardiovascular system. Zhong Yao Cai 1998;21:515-6.
- 117. Lakshmi V, Kapil RS, Patnaik GK. Spasmolytic activity of *Toddalia asiatica* var obtusifolia. Indian Drugs 1989;26:706-7.
- 118. Suanarunsawat T, Na Ayutthaya WD, Songsak T, Thirawarapan S, Poungshompoo S. Lipid-lowering and antioxidative activities of aqueous extracts of Ocimum sanctum L. leaves in rats fed with a high-cholesterol diet. Oxid Med Cell Longev 2011;2011:962025.
- 119. Cohen MM. Tulsi Ocimum sanctum: A herb for all reasons J Ayurveda Integr Med 2014;5:251-9.
- 120. Kavitha S, John F, Indira M. Amelioration of inflammation by phenolic rich methanolic extract of *Ocimum sanctum* Linn. Leaves in isoproterenol induced myocardial infarction. Indian J Exp Biol 2015;53:632-40.
- 121. Sarkar D, Srimany A, Pradeep T. Rapid identification of molecular changes in Tulsi (*Ocimum sanctum* Linn) upon ageing using leaf spray ionization mass spectrometry. Analyst 2012;137:4559-63.
- 122. Hui-Chun X, Jun S. Study on the extraction process of total anthraquinones in radix at radix et rhizoma rhei and their antilipemic effects. Afr J Tradit Complement Altern Med 2014;11:358-362.
- 123. Dludla PV, Joubert E, Muller CJ, Louw J, Johnson R. Hyperglycemia-induced oxidative stress and heart disease-cardioprotective effects of rooibos flavonoids and phenylpyruvic acid-2-O-β-D-glucoside. Nutr Metab (Lond) 2017;14:45.
- 124. Moselhy SS, Kamal IH, Kumosani TA, Huwait EA. Possible inhibition of hydroxy methyl glutaryl CoA reductase activity by nicotinic acid and ergosterol: As targeting for hypocholesterolemic action. Afr Health Sci 2016:16:319-24.
- 125. Laskarina-Maria K, Agrogiannis G, Pantopoulou A, Vlachos IS, Iliopoulos D, Karatzas T, et al. Comparative antilipidemic effect of N-acetylcysteine and sesame oil administration in diet-induced hypercholesterolemic mice. Lipids Health Dis 2010;9:23.
- 126. Kwon YS, Kim CM. Antioxidant constituents from the stem of *Sorghum bicolor*. Arch Pharm Res 2003;26:535-9.
- 127. Park KY, Jung GO, Choi J, Lee KT, Park HJ. Potent antimutagenic and their anti-lipid peroxidative effect of kaikasaponin III and tectorigenin from the flower of

- Pueraria thunbergiana. J Nat Prod 2002;65:745-7.
- 128. Butt MS, Shahzadi N, Sharif MK, Nasir M. Guar gum: A miracle therapy for hypercholesterolemia, hyperglycemia and obesity. Crit Rev Food Sci Nutr 2007;47:389-96.
- 129. Saddiqe Z, Maimoona A, Abbas G, Naeem I, Shahzad M. Pharmacological screening of *Hypericum* androsaemum extracts for antioxidant, anti-lipid peroxidation, antiglycation and cytotoxicity activity. Pak J Pharm Sci 2016 29:397-405.
- 130. Khan AL, Ali L, Hussain J, Rizvi TS, Al-Harrasi A, Lee IJ. Enzyme inhibitory radicinol derivative from endophytic fungus *Bipolaris sorokiniana* LK12, associated with *Rhazya stricta*. Molecules 2015;20:12198-208.
- 131. Khan AW, Kotta S, Ansari SH, Sharma RK, Ali J. Enhanced dissolution and bioavailability of grapefruit flavonoid Naringenin by solid dispersion utilizing fourth generation carrier. Drug Dev Ind Pharm 2015;41:772-9.
- 132. Nakchat O, Meksuriyen D, Pongsamart S. Antioxidant and anti-lipid peroxidation activities of *Tamarindus indica* seed coat in human fibroblast cells. Indian J Exp Biol 2014;52:125-32.
- 133. Chatuphonprasert W, Sangkawat T, Nemoto N, Jarukamjorn K. Suppression of beta-naphthoflavone induced CYP1A expression and lipid-peroxidation by berberine. Fitoterapia 2011;82:889-95.
- 134. Kim JI, Kim HH, Kim S, Lee KT, Ham IH, Whang WK. Antioxidative compounds from *Quercus salicina* Blume stem. Arch Pharm Res 2008;31:274-8.
- 135. Shamala S, Gunasekaran B, Shaharuddin NA, Ahmad SA, Shukor MY. Antiartherosclerotic effects of plant flavonoids. Biomed Res Int 2014;2014:480258.
- 136. Hussain J, Ali L, Khan AL, Rehman NU, Jabeen F, Kim JS, *et al.* Isolation and bioactivities of the flavonoids morin and morin-3-O-β-D-glucopyranoside from *Acridocarpus orientalis* a wild Arabian medicinal plant. Molecules 2014;19:17763-72.
- 137. Boskabady MH, Alitaneh S, Alavinezhad A. *Carum copticum* L.: A herbal medicine with various pharmacological effects. Biomed Res Int 2014;2014;569087.
- 138. Wong CP, Kaneda T, Morita H. Plant natural products as an anti-lipid droplets accumulation agent. J Nat Med 2014;68:253-66.
- 139. Suresh K, Manoharan S, Vijayaanand MA, Sugunadevi G. Chemopreventive and antioxidant efficacy of (6)-paradol in 7,12-dimethylbenz(a) anthracene induced hamster buccal pouch carcinogenesis. Pharmacol Rep 2010;62:1178-85.
- 140. Ha DT, Kim H, Thuong PT, Ngoc TM, Lee I, Hung ND, et al. Antioxidant and lipoxygenase inhibitory activity of oligostilbenes from the leaf and stem of *Vitis amurensis*. J Ethnopharmacol 2009;125:304-9.
- 141. Li GG, Bian GX, Ren JP, Wen LQ, Zhang M, Lü QJ. Protective effect of madecassoside against reperfusion

- injury after regional ischemia in rabbit heart *in vivo*. Yao Xue Xue Bao 2007;42:475-80.
- 142. Lin PJ, Chang CH. Endothelium dysfunction in cardiovascular diseases. Changgeng Yi Xue Za Zhi 1994;17:198-210.
- 143. Ng LT, Wu SJ, Tsai JY, Lai MN. Antioxidant activities of cultured *Armillariella mellea*. Prikl Biokhim Mikrobiol 2007;43:495-500.
- 144. Bian GX, Li GG, Yang Y, Liu RT, Ren JP, Wen LQ, *et al.* Madecassoside reduces ischemia-reperfusion injury on regional ischemia induced heart infarction in rat. Biol Pharm Bull 2008;31:458-63.
- 145. Fernando CD, Soysa P. Total phenolic, flavonoid contents, *in-vitro* antioxidant activities and hepatoprotective effect of aqueous leaf extract of *Atalantia ceylanica*. BMC Complement Altern Med 2014:14:395.
- 146. Nattharat J, Chatuphonprasert W, Miroestrol KJ. A phytoestrogen from *Pueraria mirifica*, improves the antioxidation state in the livers and uteri of β-naphthoflavone-treated mice. J Nat Med 2014:68:173-80.
- 147. Lin YL, Chang YY, Kuo YH, Shiao MS. Anti-lipid-peroxidative principles from *Tournefortia sarmentosa*. J Nat Prod 2002;65:745-7.
- 148. Cheng HY, Lin TC, Yu KH, Yang CM, Lin CC. Antioxidant and free radical scavenging activities of *Terminalia chebula*. Biol Pharm Bull 2003;26:1331-5.
- 149. Choi L, Han YN, Lee KT, Park KY, Kwak TS, Kwon SH, *et al.* Anti-lipid peroxidative principles from the stem bark of *Kalopanax pictus* Nakai. Arch Pharm Res 2001;24:536-40.
- 150. Janicsák G, Hohmann J, Zupkó I, Forgo P, Rédei D, Falkay G, *et al.* Diterpenes from the aerial parts of *Salvia candelabrum* and their protective effects against lipid peroxidation. Planta Med 2003;69:1156-9.
- 151. Manandhar B, Paudel KR, Sharma B, Karki R. Phytochemical profile and pharmacological activity of *Aegle marmelos* Linn. J Integr Med 2018;16:153-63.
- 152. Dhiman AK. In: Discussion of Plants, Sacred Plants and their Medicinal Uses. New Delhi: Daya Publication House; 2003. p. 18-9.
- 153. Parmar C, Kaushal MK. Aegle marmelos correa. In: Wild Fruits of the Sub-Himalayan Region. New Delhi: Kalyani Publishers; 1982. p. 1-5.
- 154. Purohit SS, Vyas SP. In: *Aegle marmelos* Correa ex Roxb. (Bael), Medicinal Plant Cultivation a Scientific Approach. Jodhpur: Agrobios; 2004. p. 280-5.
- 155. Manik S, Gauttam V, Kalia AN. Antidiabetic and antihyperlipidemic effect of allopolyherbal formulation in OGTT and STZ-induced diabetic rat model. Indian J Exp Biol 2013;51:702-8.
- 156. Maity P, Hansda D, Bandyopadhyay U, Mishra DK. Biological activities of crude extracts and chemical constituents of Bael, *Aegle marmelos* (L.) Corr. Indian J Exp Biol 2009;47:849-61.
- 157. Sangweni NF, Dludla PV, Mosa RA, Kappo AP,

- Opoku A, Muller CJ, et al. Lanosteryl triterpenes from *Protorhus longifolia* as a cardioprotective agent: A mini review. Heart Fail Rev 2019;24:155-66.
- 158. Panda S, Kar A. Periplogenin-3-O- -D-glucopyranosyl -(1->6)- -D-glucopyranosyl- -(1->4) -D-cymaropyranoside, isolated from *Aegle marmelos* protects doxorubicin induced cardiovascular problems and hepatotoxicity in rats. Cardiovasc Ther 2009;27:108-16.
- 159. Ginter E, Simko V. Plant polyphenols in prevention of heart disease. Bratisl Lek Listy 2012;113:476-80.
- 160. Chan JY, Yuen AC, Chan RY, Chan SW. A review of the cardiovascular benefits and antioxidant properties of allicin. Phytother Res 2013;27:637-46.
- 161. Slevin M, Ahmed N, Wang Q, McDowell G, Badimon L. Unique vascular protective properties of natural products: Supplements or future main-line drugs with significant anti-atherosclerotic potential? Vasc Cell 2012;4:9.
- 162. Cavagnaro PF, Galmarini CR. Effect of processing and cooking conditions on onion (*Allium cepa* L.) induced antiplatelet activity and thiosulfinate content. J Agric Food Chem 2012;60:8731-7.
- 163. Arora RC, Arora S, Gupta RK. The long-term use of garlic in ischemic heart disease An appraisal. Atherosclerosis 1981;40:175-9.
- 164. Wohlrab J, Wohlrab D, Marsch WC. Acute effect of a dried ethanol-water extract of garlic on the microhaemovascular system of the skin. Arzneimittelforschung 2000;50:606-12.
- Wolf S, Reim M, Jung F. Effect of garlic on conjunctival vessels: A randomised, placebo-controlled, doubleblind trial. Br J Clin Pract Suppl 1990;69:36-9.
- 166. Ashraf MZ, Hussain ME, Fahim M. Endothelium mediated vasorelaxant response of garlic in isolated rat aorta: Role of nitric oxide. J Ethnopharmacol 2004;90:5-9.
- 167. (a). Jung EM, Jung F, Mrowietz C, Kiesewetter H, Pindur G, Wenzel E. Influence of garlic powder on cutaneous microcirculation. A randomized placebocontrolled double-blind cross-over study in apparently healthy subjects. Arzneimittelforschung 1991;41:626-30. (b). Jung F, Jung EM, Mrowietz C, Kiesewetter H, Wenzel E. Influence of garlic powder on cutaneous microcirculation. A randomized placebo-controlled double-blind cross-over study in apparently healthy subjects. The effects of garlic powder on cutaneous microcirculation. A cross-over test with healthy test persons. Br J Clin Pract Suppl 1990;69:30-5.
- 168. Breithaupt-Grögler K, Ling M, Boudoulas H, Belz GG. Protective effect of chronic garlic intake on elastic properties of aorta in the elderly. Circulation 1997;96:2649-55.
- 169. Das I, Khan NS, Sooranna SR. Potent activation of nitric oxide synthase by garlic: A basis for its therapeutic applications. Curr Med Res Opin 1995;13:257-63.
- 170. Dirsch VM, Kiemer AK, Wagner H, Vollmar AM.

- Effect of allicin and ajoene, two compounds of garlic, on inducible nitric oxide synthase. Atherosclerosis 1998;139:333-9.
- 171. Pedraza-Chaverri J, Tapia E, Medina-Campos O, de los Angeles Granados M, Franco M. Garlic prevents hypertension induced by chronic inhibition of nitric oxide synthesis. Life Sci 1998;62:71-7.
- 172. Ku DD, Abdel-Razek TT, Dai J, Fallon MB, Abrams GA. Mechanisms of garlic induced pulmonary vasorelaxation: Role of allicin. Circulation 1997;96:106.
- 173. Baluchnejadmojarad T, Roghani M, Homayounfar H, Hosseini M. Beneficial effect of aqueous garlic extract on the vascular reactivity of streptozotocin-diabetic rats. J Ethnopharmacol 2003;85:139-44.
- 174. Baluchnejadmojarad T, Roghani M. Endothelium-dependent and independent effect of aqueous extract of garlic on vascular reactivity on diabetic rats. Fitoterapia 2003;74:630-7.
- 175. Block E, Purcell PF, Yolen SR. Onions and heartburn. Am J Gastroenterol 1992;87:679-80.
- 176. Majewska-Wierzbicka M, Czeczot H. Flavonoids in the prevention and treatment of cardiovascular diseases. Pol Merkur Lekarski 2012;32:50-4.
- 177. Seref A, Aktas C, Uygur R, Topcu B, Kanter M, Erboga M, *et al.* Antioxidant and anti-apoptotic effects of onion (*Allium cepa*) extract on doxorubicin-induced cardiotoxicity in rats. J Appl Toxicol 2013;33:202-8.
- 178. Brüll V, Burak C, Stoffel-Wagner B, Wolffram S, Nickenig G, Müller C, *et al.* Effects of a quercetinrich onion skin extract on 24 h ambulatory blood pressure and endothelial function in overweight-to-obese patients with (pre-)hypertension: A randomised double-blinded placebo-controlled cross-over trial. Br J Nutr 2015;114:1263-77.
- 179. Ro JY, Ryu JH, Park HJ, Cho HJ. Onion (*Allium cepa* L.) peel extract has anti-platelet effects in rat platelets. Springerplus 2015;4:17.
- 180. Boligon AA, Pimentel VC, Bagatini MD, Athayde ML. Effect of *Scutia buxifolia* Reissek in nucleotidase activities and inhibition of platelet aggregation. J Nat Med 2015;69:46-54.
- 181. Mosawy S, Jackson DE, Woodman OL, Linden MD. The flavonols quercetin and 3', 4'-dihydroxyflavonol reduce platelet function and delay thrombus formation in a model of Type 1 diabetes. Diab Vasc Dis Res 2014;11:174-81.
- 182. Lee SM, Moon J, Chung JH, Cha YJ, Shin MJ. Effect of quercetin-rich onion peel extracts on arterial thrombosis in rats. Food Chem Toxicol 2013;57:99-105.
- 183. Hubbard GP, Wolffram S, de Vos R, Bovy A, Gibbins JM, Lovegrove JA. Ingestion of onion soup high in quercetin inhibits platelet aggregation and essential components of the collagen-stimulated platelet activation pathway in man: A pilot study. Br J Nutr 2006;96:482-8.
- 184. Vilahur G, Badimon L. Antiplatelet properties of

- natural products. Vasc Pharmacol 2013;59:67-75.
- 185. Singh SK, Suresh MV, Voleti B, Agrawal A. The connection between C-reactive protein and atherosclerosis. Ann Med 2008;40:110-20.
- 186. Nadia A, Ur-Rehman K, Jahan N, Ijaz M, Manzoor A, Khan KM, et al. Cardioprotective and metabolomic profiling of selected medicinal plants against oxidative stress. Oxid Med Cell Longev 2018;2018:98.
- 187. Li XS, Chen GR, Li JM, Hu Y, Wang F, Hu Y. The beneficial effect of *Gingko biloba* extract on myocardial impairment in diabetic rats. Zhongguo Ying Yong Sheng Li Xue Za Zhi 2005;21:176-8.
- 188. Zhang YN, Zhang J, Huang GQ, Gu CY, Chen WZ. Protective effects of extract of *Ginkgo biloba* (EGb) on lysophosphatidylcholine induced damages of vascular endothelial cells *in vitro*. Yao Xue Xue Bao 1997;32:735-9.
- 189. Dhalwal K, Deshpande YS, Purohit AP. Evaluation of *in vitro* antioxidant activity of *Sida rhombifolia* (L.) ssp. *retusa* (L.). J Med Food 2007;10:683-8.
- 190. Wu L, Qiao H, Li Y, Li L. Protective roles of puerarin and Danshensu on acute ischemic myocardial injury in rats. Phytomedicine 2007;14:652-8.
- 191. Shah R, Gulati V, Palombo EA. Pharmacological properties of guggulsterones, the major active components of gum guggul. Phytother Res 2012;26:1594-605.
- 192. Verma, N. Introduction to hylerlipidemia and its treatment: A review. Int J Curr Pharm Res 2016;9:6-14.
- 193. Shelke S, Khairnar A, Rathod V, Kalawane Y, Jagtap A.

- Reviw on antihyperlipedemia lipophilic drugs and their novel formulation approaches. Int J Pharm Pharm Sci 2017;9:1-8.
- 194. Thomas L, Kamath JV. Evaluation of antihyperlipidemic activity of *Capsicum frutiescens*. Int J Curr Pharm Res 2017;9:165-8.
- 195. Chakrobarty U, Das H. Antidiabetic and antioxidant activities of Cinnamomum tamala leaf extract in Stz-treated diabetic rat. Glob J Biotech Biochem 2010;5:12-8.
- 196. Bajaj S, Srinivasan BP. Investigations into the antidiabetic activity of Azadirachta indica. Scoparia dulcis. Indian J Pharmacol 1998;31:138-41.
- 197. Chandola HM, Tripathi SN, Udupa KN. Effect of Cinnamomum tamala on plasma insulin vis-à-vis blood sugar in patients of 269 diabetes mellitus. J Res Ayur Sidha 1980;1:345-57.
- 198. Tuzcu Z, Orhan C, Sahin N, Juturu V, Sahin K. Cinnamon polyphenol extract inhibits hyperlipidemia and inflammation by modulation of transcription factors in high-fat diet-fed rats. Oxid Med Cell Longev 2017;2017:1583098.
- 199. Kim GJ, Lee JY, Choi HG, Kim SY, Kim E, Shim SH, et al. Cinnamomulactone, a new butyrolactone from the twigs of *Cinnamomum cassia* and its inhibitory activity of matrix metalloproteinases. Arch Pharm Res 2017;40:304-10.

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